

Virtual Ligand Screening Combined with NMR to Identify Dvl PDZ Domain Inhibitors Targeting the Wnt Signaling

Jufang Shan and Jie J. Zheng

Abstract

Virtual ligand screening is a powerful technique to identify potential hits of targets and to increase hit rates. Here, we describe how we used this technique combined with NMR ^{15}N HSQC experiments to obtain small molecules that bind to the PDZ domain of Dvl targeting the Wnt signaling pathway.

Key words: Virtual ligand screening, Database search, Docking, NMR, ^{15}N HSQC, SAR

1. Introduction

Wnt signaling pathways play key developmental and growth regulatory roles that significantly affect many biological processes; abnormal Wnt activity has been implicated in cancer and other human diseases. Dishevelled (Dvl) relays the signal from the membrane-bound Wnt receptors of the Frizzled (Fz) family to downstream substrates via its PDZ domain (1). The special role of the Dvl PDZ domain in the Wnt signaling pathway (1) makes it a potential pharmaceutical target (2). Small organic inhibitors of the PDZ domain in Dvl (3, 4) might be useful in dissecting molecular mechanisms and formulating pharmaceutical agents that target cancers or other diseases in which Wnt signaling is involved.

1.1. Structure-Based Virtual Ligand Screening for Inhibitor of the Dvl PDZ Domain

Like many other PDZ domains (5), the structure of the Dvl PDZ domain is known (6). This has permitted us to use structure-based virtual ligand screening (VLS) to computationally access potential ligands. By using a UNITY search for compounds with the potential to bind to the PDZ domain, FlexX docking of candidates into the binding site, Cscore ranking of binding modes, and chemical shift perturbation NMR experiments, we identified a small organic

molecule (NSC668036) from the National Cancer Institute (NCI) small-molecule library that can bind to the mDvl1 PDZ domain. Further NMR experiments confirmed that the compound binds to the peptide-binding site on the surface of the PDZ domain. In addition, we carried out molecular dynamics (MD) simulations of the interaction between this compound and the PDZ domain as well as that between the C-terminal region of a known PDZ domain inhibitor (Dapper) and the PDZ domain, and we compared the binding free energies of these interactions calculated via the molecular mechanics Poisson–Boltzmann surface area (MM-PBSA) method (7–9).

1.2. Optimizing Dvl PDZ Domain Inhibitor by Virtually Exploring Chemical Space

With the advancement of technology in the field of drug discovery, hits of a potential therapeutic reagent can be identified in a comparatively straightforward fashion by using high-throughput screening (10, 11). However the follow-up hit-to-lead process and lead optimization still remain as challenging problems in the drug discovery process (12, 13). One of the most frequently taken approaches in the hit-to-lead process is hit evolution (13). During hit evolution, analogues of the most promising hits are synthesized for the development of structure–activity relationship (SAR) data. The SAR is then used to guide the synthesis and optimization of lead compounds to improve their potencies and physicochemical properties, and to reduce off-target activities. The synthesis processes are usually long and labor intensive. Virtual screening of databases consisting of physically available compounds may help us to take advantage of the chemistry that has already been done and speed up projects, especially with the ever-growing list of existing compounds. Indeed, the Zinc database has 13 million compounds (14) and the iResearch™ Library (ChemNavigator, San Diego, CA) has more than 50 million unique chemicals. Although the databases of available compounds are still under-sampled (15), the chemical space represented by those millions of compounds should never be neglected. We believed that the large chemical space of available compounds offers us with an opportunity to explore SAR of known hits; and as a proof of principle test, we searched the ChemDiv database for the Dvl PDZ domain inhibitors based on an inhibitor identified above (16). In our studies, we first developed a pharmacophore model based on NSC668036; based on the model, we then screened the ChemDiv database by using an algorithm that combines similarity search and docking procedures; finally, we selected potent inhibitors based on docking analysis and examined them by using NMR spectroscopy. NMR experiments showed that all the 15 compounds we chose bound to the PDZ domain tighter than NSC668036.

2. Materials

2.1. Virtual Screening

1. The structure of the Dvl PDZ domain was obtained from the protein data bank (PDB entry 1L6O (6)) (Note 1).
2. The three-dimensional (3D) NCI database of small-molecule is available from NCI at no cost, and it includes the coordinates of more than 250,000 drug-like chemical compounds. The Tripos format of NCI database comes with SYBYL® (St Louis, MO) and is ready for UNITY search. For ChemDiv database, UNITY in SYBYL® was used to convert the SLN files obtained from ChemDiv Inc. (San Diego, CA).

2.2. NMR Experiments

1. The ¹⁵N-labeled mouse Dvl1 PDZ domain (residues 247–341 of mouse Dvl1) was prepared as described previously (1, 17) by the protein production facility at St. Jude Children's Research Hospital. For NMR, the 0.3 mM ¹⁵N-labeled Dvl PDZ domain was in 100 mM potassium phosphate buffer (pH 7.5), 10% D₂O, and 0.5 mM EDTA.
2. Compounds from the NCI library were obtained from Drug Synthesis and Chemistry Branch, Developmental Therapeutics Program, Division of Cancer Treatment and Diagnosis, NCI (<http://129.43.27.140/ncidb2/>). The 15 ChemDiv compounds were purchased from ChemDiv Inc. (San Diego, CA). Compounds were dissolved in the 100 mM potassium phosphate buffer (pH 7.5) with 10% D₂O, and 0.5 mM EDTA. DMSO was added for compounds not soluble in the above buffer (see Note 3).

3. Methods

3.1. Structure-Based Ligand Screening of Small Compounds Binding to the PDZ Domain

1. The UNITY module of the SYBYL software package (Tripos, Inc.) was used to screen the NCI small-molecule three-dimensional database for chemical compounds that could fit into the peptide-binding groove of the Dvl PDZ domain. Based on the complex structure of PDZ domain and Dapper peptide (6), a search query was designed; it consists of two hydrogen bond donors (backbone amide nitrogens of Gly266 and Ile269) and two hydrogen bond acceptors (carbonyl oxygens of Ile267 and Ile269) on the PDZ domain, with 0.3 Å tolerances for spatial constraints (Fig. 1).
2. The Flex search module of UNITY was used to explore the 3D small-molecule NCI database to identify compounds that met the requirements of the query. The Flex search option of

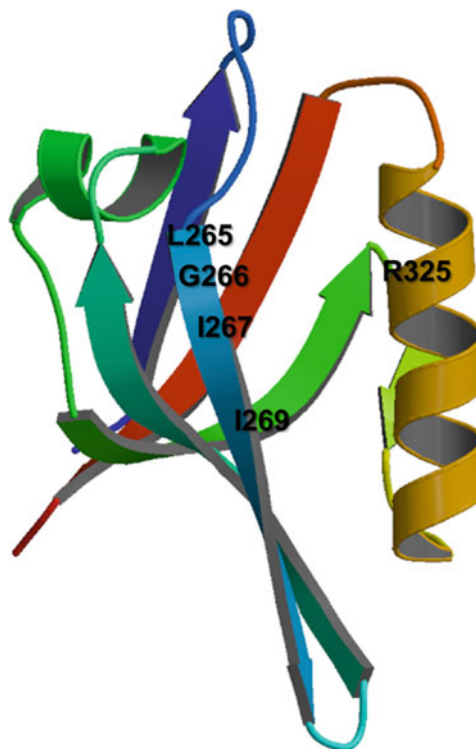


Fig. 1. The structure of the Dvl PDZ domain. The ligand site is located at α -helix B (yellow) and β -sheet B (light blue) regions.

UNITY considers the flexibility of compounds, and it uses the Directed Tweak algorithm to conduct a rapid and conformationally flexible 3D search (18). The search yielded 108 organic compounds as the initial hits.

3. These 108 hits then were “docked” into the binding site of the PDZ domain by using the FlexX program of SYBYL. FlexX is energy minimization modeling software that varies the conformation of the ligand to fit it into the protein-binding site (19). The receptor’s binding site was defined by residues Gly266, Ile269, and Arg325 with a selection radius of 5.9 Å, and a core subpocket was defined by Gly266 with a selection radius of 5.9 Å (Fig. 1). The docking condition was validated by docking the dapper peptide to the PDZ domain as in Note 2.
4. The results of the docking procedure were evaluated manually and those compounds that were not docked into the binding pocket of the PDZ domain were removed. The Cscore program of SYBYL was then used to rank the remaining compounds on the basis of their predicted ability to bind to the binding pocket. Cscore generates a relative, consensus score, based on the individual scoring functions of the protein–ligand complex (20). One of the scoring functions in Cscore, the

F_{score}, is particularly useful. F_{score} considers polar and nonpolar interactions in calculating the binding free energy of ligand and protein. Based on this notion, nine compounds whose F_{score}s were better than that of the control Dapper–PDZ interaction were then further characterized and subsequently obtained from the Developmental Therapeutics Program of the NCI.

3.2. Testing VLS Hits with NMR Spectroscopy

The abilities of the nine compounds obtained from DTP to bind to the PDZ domain were tested by NMR spectroscopy, mainly the chemical shift perturbation experiment (21). NMR ¹⁵N HSQC experiments were performed by using a Varian Inova 600 MHz NMR spectrometer at 25°C.

1. HSQC of the ¹⁵N-labeled Dvl PDZ domain (residues 247–341 of mouse Dvl1) was recorded as a reference.
2. Compound was added to the solution PDZ domain and NMR spectrum of the mixture was recorded. If the compound generated perturbations to the PDZ domain, different amounts of compound were added to the PDZ domain and series of NMR spectra were recorded for calculations of compound's dissociation constant (K_D) to the PDZ domain.
3. NMR spectra were processed with NMRpipe (22).
4. NMR spectra were analyzed by using Sparky (23). We monitored whether added compounds generated chemical shift perturbations to the resonances of the Dvl PDZ domain and compared if the chemical shift perturbations were similar to those caused by binding of the Dapper peptide and Fz7 peptide, which was derived from an Fz membrane receptor (1). Similar perturbations would suggest that compound binds to the same binding site as native PDZ domain-binding partners such as Dapper and Fz. Among nine tested compounds, NSC668036 perturbed the PDZ domain similarly as Dapper and Fz suggesting that it binds to the same binding site as these two native peptides (Note 4).
5. Calculating K_D by using HSQC titration spectra: The binding affinities (K_D) of PDZ ligands were calculated using HSQC spectra by following the method described by Worrall et al. (24). The mean chemical-shift perturbation changes caused by the binding of ligands were calculated using Eq. 1. K_D was then calculated using Eqs. 2 and 3 by applying a one-site binding model with corrections for dilutions, where R was the ligand-to-protein molar ratio, P was the protein concentration before titration, C was the ligand stock concentration, and K_D was the dissociation constant. Two-parameter nonlinear least-squares fitting was performed with program Prism (GraphPad Software, La Jolla, CA):

$$\Delta\delta_{\text{avg}} = \sqrt{1/2(\Delta\delta N/5)^2 + (\Delta\delta H)^2} \quad (1)$$

$$\Delta\delta_{\text{binding}} = (1/2)\Delta\delta_{\text{max}}(A - \sqrt{A^2 - 4R}). \quad (2)$$

$$A = 1 + R + (PR + C)K_D / (PC). \quad (3)$$

3.3. Molecular Dynamics Simulation of Verified PDZ Ligand

To further investigate the interaction between the PDZ domain and NSC668036, the AMBER software (25) suite was used to conduct an MD simulation study of the NSC668036–PDZ domain complex.

1. The starting structures of ligand–protein complexes were prepared using the output from the FlexX docking studies. To sample sufficient possible binding modes during the MD simulation, the entire output of the initial FlexX docking results was reexamined. Based on structure alignment, the 30 docked NSC668036 conformers can be grouped into three clusters. Manual inspection of these docking conformers led us to select 10 conformers from the three clusters representing the 30 conformers as starting points for the MD simulations.
2. After neutralization of these complexes with Na^+ or Cl^- , they were dissolved in a periodic rectangular TIP3P water box, with each side 10 Å from the edge of the complex. AM1-BCC charges and parameters from GAFF force field were assigned to NSC668036 by using the Antechamber module (26) in AMBER 8. For protein, ions, and water, parm99 force field (25, 27) was used.
3. Systems were minimized by a 1,000-step steepest descent minimization followed by a 9,000-step conjugated gradient minimization (25, 27).
4. MD simulation was performed by using the sander program in AMBER 8 with a time step of 2 fs and the nonbonded cutoff set to 9.0 Å. Constant volume (NVT) and constant pressure (NPT) ensemble simulations were carried out to equilibrate the system. In detail, a 50 ps NVT simulation was used to increase the temperature from 100 to 300 K; then the 50 ps NPT ensemble was used to adjust the solvent density, and another 100 ps NPT ensemble was used to gradually reduce the harmonic restraints from 5.0 kcal/mol/Å² to none (25, 27).
5. MD simulations were performed in explicit water for 5 ns after equilibration with the particle mesh Ewald (PME) method (28, 29) using the NPT ensemble. During the production run, snapshots were saved every 5 ps. Other simulation parameters were set to values similar to those described in the work by Gohlke et al. (30).

3.4. Binding Free Energy Calculation

1. The MM-PBSA algorithm (7–9) was then used to calculate the binding free energy of the interaction between the PDZ domain and NSC668036 using Eq. 4 with the mm_pbsa.pl script in AMBER 8, which employs an MM-PBSA approach:

$$\Delta G_{\text{total}} = G^{\text{complex}} - G^{\text{protein}} - G^{\text{ligand}}, \quad (4)$$

where

$$G = H_{\text{gas}} + H_{\text{trans/rot}} + G_{\text{solvation}} - TS \quad (5)$$

$$G_{\text{solvation}} = G_{\text{solvation}}^{\text{polar}} + G_{\text{solvation}}^{\text{nonpolar}} \quad (6)$$

$$G_{\text{solvation}}^{\text{nonpolar}} = \gamma A + b, \quad (7)$$

where the gas phase energy, H_{gas} , is the sum of internal (bond, angle, and torsion), van der Waals, and electrostatic energies in the molecular mechanical force field with no cutoff, as calculated by molecular mechanics (31). $H_{\text{trans/rot}}$ is $3RT$ (R being the gas constant) because of six translational and rotational degrees of freedom. The solvation free energy, $G_{\text{solvation}}$, was calculated by using the PB model (8, 30, 31). In PB calculations, the polar solvation energy, $G_{\text{solvation}}^{\text{polar}}$, was obtained by solving the PB equation with Delphi using parse radius, parm94 charges (for the PDZ domain and the Dapper peptide), and AM1-BCC charges (for the compound). The nonpolar contribution was calculated by Eq. 7. In this equation, A is the solvent accessible area calculated by the Molsurf module in Amber 8 and γ (surface tension) and b (a constant) were 0.00542 kcal/mol/Å² and 0.92 kcal/mol, respectively.

2. All of the energy terms given above were averaged from 150 snapshots extracted every 20 ps, and the entropy TS was estimated by normal-mode analysis using 15 snapshots extracted every 200 ps during the last 3-ns production run.
3. During the 10 MD simulation runs, the simulation that started with conformer 22 had the lowest and most stable binding free energy, suggesting that this conformer represents the true PDZ domain-bound conformation of NSC668036 in solution.

3.5. Deduction of Pharmacophore Based on a PDZ-Inhibitor Complex Structure and Two Non-binders

A pharmacophore is composed of functional groups essential and necessary for receptor–ligand binding; without those groups, ligands will no longer bind to receptors and lose their activities (32). Pharmacophore-based approaches have been widely used and shown successes in the field of computer-aided drug design (33, 34). In order to identify more PDZ domain inhibitors, we took such an approach, derived the pharmacophore of PDZ ligands, and used it to screen for PDZ domain inhibitors in the ChemDiv database.

1. The complex structure of NSC668036 and the PDZ domain generated by docking and extensive molecular dynamics simulations was used to build a pharmacophore by using LigandScout (Inte:Ligand, Austria). LigandScout extracts 3D pharmacophores based on complex structures (35).
2. The LigandScout pharmacophore model of NSC668036 is consistent with the structural analysis. The carboxyl group contributes three hydrogen-bond acceptors and the isopropyl group contributes a hydrophobic interaction to the pharmacophore.
3. The NSC668036 compound was compared with other compounds that did not bind to the PDZ domain as verified by NMR experiments. By aligning two non-binders, NSC344681 ((((2-amino-3-hydroxybutanoyl) amino)acetyl)amino)acetic acid) and NSC119132 (methyl 3-((2-amino-3-(aminooxy) propanoyl)amino)-2-((aminooxy) methyl)-3-oxopropanoate) against NSC668036, the differences among these structures that might render NSC344681 and NSC119132 inactive were identified. NSC344681 does not have a hydrophobic group at the 2-position; and NSC119132 is an ester instead of a free acid. This finding strongly suggested that both the carboxyl group and the hydrophobic group next to it were important to the binding and might compose the pharmacophore of PDZ ligands since compounds without them did not bind to the protein.

3.6. Pharmacophore-Based Virtual Screening

1. Based on the pharmacophore proposed above, a 2D search to retrieve all the compounds with 2-(3-methylbutanoic acid) groups in the ChemDiv database by using UNITY in SYBYL® (Tripos, Inc.) was carried out and the search returned 116 hits.
2. To reduce the number of compounds needed to test experimentally and to select ligands with higher potentials to bind to the PDZ domain, the hits were further filtered by using FlexX docking and “Cscore” ranking following the procedure described in Subheading 2.3.1. After manually inspecting top-ranked compounds, 15 compounds were chosen for further examinations based on the following four criteria: (a) the compounds were docked into the designated binding site, (b) their docking conformations were complementary to those of the PDZ domain, and (c) the docked compounds formed hydrogen-bonds with the β A- β B loop as well as (d) additional hydrogen bonds with the PDZ domain.

3.7. Build SAR Models of PDZ Domain Inhibitors

1. The interactions between the 15 compounds and the Dvl PDZ domain were examined experimentally by using NMR chemical shift perturbation experiments as described in Subheading 2.3.2. All the 15 compounds bound to the PDZ domain at the same binding site as the Fz7 peptide.

2. The above-verified compounds were then ranked by their K_D to the PDZ domain, which were calculated by using chemical-shift perturbation changes as described in Subheading 2.3.2.
3. The complex structures of these compounds with the PDZ domain were modeled. The FlexX docking models of the 15 PDZ ligands were refined by using Glide (Schrödinger Inc.). During Glide docking, compounds' amide bonds were kept rigid; hydrogen-bond pharmacophores were designed on the protein to induce ligands to form hydrogen-bonds with the β A- β B loop and the β B strand of the PDZ domain. Other than fixed amide bonds and H-bond pharmacophores, default docking parameters were used. All ligand binding poses generated by Glide have reasonable Glide scores, suggesting that they are likely very close to the true binding modes. For example, the Glide score of compound ChemDiv 5435-0027 is -7.57 . According to Schrödinger Inc., low-micromolar inhibitors should have scores around -7 . Glide 2.5 predicted binding affinities of a set of 125 crystallized complexes with an RMSD of 2.2 kcal/mol against experimental data (36). Based on the facts that compound ChemDiv 5435-0027 binds to the PDZ domain with a moderate binding affinity and its glide score fits with the experimental data, it is likely that this docking conformation is close to the true binding mode. This conformation was then used to generate complex structures for compounds ChemDiv 2509-0036 and 2509-0040 which had different docking poses than those of the rest of 13 compounds. Complex structures of compounds ChemDiv 2509-0036 and 2509-0040 were modeled by superimposing them onto docked compound ChemDiv 5435-0027 followed by ligand minimization in the ligand-binding pocket of the PDZ domain with LigandScout.
4. Structure-based pharmacophore models of these compounds were built using LigandScout as in Subheading 2.3.5.
5. These 15 compounds are similar in structure but have considerable differences in binding affinities, and thus provide an opportunity to study their SAR to gain insights into the molecular determinants of PDZ-ligand binding which might facilitate further ligand optimization. These 15 compounds have similar scaffolds except that some of them have an extra double bond in their scaffold. According to their scaffolds, these compounds can be grouped into two classes and by comparing their structures and binding strengths in the context of docking complex structures and complex structure-based pharmacophore model, the SAR models of these 15 compounds can be built. In turn, the SAR models can be used to guide further optimization of PDZ ligands.

4. Notes

1. Using the Structure of a Homologous Protein in VLS and Docking

To build the search query for the virtual screening stage, we used the crystal structure of the PDZ domain of *Xenopus* Dvl bound with the Dapper peptide (6) instead of the NMR solution structure of the apo-PDZ domain of mouse Dvl (1). The two PDZ domains share a high degree of homology, especially around the peptide-binding sites; near the binding sites, there is only a single amino acid difference between the two PDZ domains (Glu323 in the PDZ domain of mDvl1 vs. Asp326 in the PDZ domain of *Xenopus* Dvl), and the side chain of this residue points away from the peptide-binding cleft. The peptide-binding cavity of the domain is smaller in the apo form of the solution structure than in the crystal structure of the Dapper-bound PDZ domain of *Xenopus* Dvl. This difference is consistent with the classic “induce-and-fit” mechanism, in which, upon the binding of a peptide or a small organic molecule, the binding sites in the PDZ domain undergo conformational change to accommodate the bound ligand. However, this flexibility cannot be fully explored through a UNITY search and the FlexX docking protocols. Therefore, although the PDZ domain of mouse Dvl was used in the experimental studies, the crystal structure of the PDZ domain of *Xenopus* Dvl provides a better template for the virtual screening steps. Indeed, the binding free energies calculated from MD simulation of the PDZ domain–NSC668036 and PDZ domain–Dapper peptide complexes fit well with the experimental binding data.

2. Docking Validation

As a control, we also docked the Dapper peptide into the PDZ domain using FlexX. Under this condition, the docked Dapper peptide had a conformation similar to that found in the crystal structure of the complex with a backbone RMSD of 2.04 Å. In particular, the backbone RMSD for the last six C-terminal amino acids is 1.22 Å, indicating that the docking procedure we used was able to dock ligand into the binding site of the PDZ domain with reasonable accuracy.

3. Compound Handling

To test compounds using NMR experiments, they should be dissolved in the same buffer as in which the protein was dissolved. pH should be adjusted back to 7.5 if changed to ensure that perturbations on the PDZ domain would only come from the compounds but not from changes in the condition. For compounds with small solubility in the protein buffer, small amount of DMSO was added so that there would be no more than 5%

of DMSO in the final mixture of the PDZ domain with the compound. NMR experiments showed that <5% DMSO does not change the spectra of the PDZ domain. Compound solutions were stored in the freezer at -20°C to avoid hydrolysis and/or other reactions that might change compound identities.

4. Test Selectivity of Identified Ligands

We tested two other PDZ domains: the first PDZ domain of PSD-95, PSD95a (37) (PDB entries 1IU0 and 1IU2), which belongs to the class I PDZ domains, and the PDZ7 domain of the glutamate receptor-interacting protein (38) (PDB entry 1M5Z), a member of the class II PDZ domains. NSC668036 binds to both of these PDZ domains extremely weakly.

References

1. Wong H-C, et al. Direct binding of the PDZ domain of Dishevelled to a conserved internal sequence in the c-terminal region of Frizzled. *Mol Cell*. 2003;12:1251–60.
2. Barker N, Clevers H. Mining the Wnt pathway for cancer therapeutics. *Nat Rev Drug Discov*. 2006;5:997–1014.
3. Grandy D, et al. Discovery and characterization of a small molecule inhibitor of the PDZ domain of dishevelled. *J Biol Chem*. 2009;284:16256–63.
4. Lee HJ, et al. Sulindac inhibits canonical Wnt signaling by blocking the PDZ domain of the protein Dishevelled. *Angew Chem Int Ed Engl*. 2009;48:6448–52.
5. Lee HJ, Zheng JJ. PDZ domains and their binding partners: structure, specificity, and modification. *Cell Commun Signal*. 2010;8:8.
6. Cheyette BNR, et al. Dapper, a Dishevelled-associated antagonist of b-Catenin and JNK signaling, is required for Notochord formation. *Dev Cell*. 2002;2:449–61.
7. Gohlke H, Case DA. Converging free energy estimates: MM-PB(GB)SA studies on the protein–protein complex Ras–Raf. *J Comput Chem*. 2004;25:238–50.
8. Wang J, et al. Use of MM-PBSA in reproducing the binding free energies to HIV-1 RT of TIBO derivatives and predicting the binding mode to HIV-1 RT of efavirenz by docking and MM-PBSA. *J Am Chem Soc*. 2001;123:5221–30.
9. Wang W, et al. BIOMOLECULAR SIMULATIONS: Recent developments in force fields, simulations of enzyme catalysis, protein–ligand, protein–protein, and protein–nucleic acid noncovalent interactions. *Annu Rev Biophys Biomol Struct*. 2001;30:211–43.
10. Burbaum JJ, Sigal NH. New technologies for high-throughput screening. *Curr Opin Chem Biol*. 1997;1:72–8.
11. Liu B, et al. Technological advances in high-throughput screening. *Am J Pharmacogenomics*. 2004;4:263–76.
12. Bleicher KH, et al. Hit and lead generation: beyond high-throughput screening. *Nat Rev Drug Discov*. 2003;2:369–78.
13. Keseru GM, Makara GM. Hit discovery and hit-to-lead approaches. *Drug Discov Today*. 2006;11:741–8.
14. Irwin JJ, Shoichet BK. ZINC—a free database of commercially available compounds for virtual screening. *J Chem Inf Model*. 2004;45:177–82.
15. Hann MM, Oprea TI. Pursuing the leadlikeness concept in pharmaceutical research. *Curr Opin Chem Biol*. 2004;8:255–63.
16. Shan J, et al. Identification of a specific inhibitor of the Dishevelled PDZ domain. *Biochemistry*. 2005;44:15495–503.
17. London TBC, et al. Interaction between the internal motif KTXXXI of Idax and mDvl PDZ domain. *Biochem Biophys Res Commun*. 2004;322:326–32.
18. Hurst T. Flexible 3D searching: the directed tweak technique. *J Chem Inf Comput Sci*. 1994;34:190–6.
19. Rarey M, et al. A fast flexible docking method using an incremental construction algorithm. *J Mol Biol*. 1996;261:470–89.
20. Clark RD, et al. Consensus scoring for ligand/protein interactions. *J Mol Graph Model*. 2002;20:281–95.
21. Zheng J, et al. Identification of the binding site for acidic phospholipids on the PH domain of

- dynammin: Implications for stimulation of GTPase activity. *J Mol Biol.* 1996;255:14–21.
22. Delaglio F, et al. NMRPipe: a multidimensional spectral processing system based on UNIX pipes. *J Biomol NMR.* 1995;6:277–93.
 23. Goddard TD, Kenlller DG SPARKY 3. University of California, San Francisco 2008.
 24. Worrall JAR, et al. Transient protein interactions studied by NMR spectroscopy: the case of cytochrome c and adrenodoxin. *Biochemistry.* 2003;42:7068–76.
 25. Case DA, et al. AMBER 8. La Jolla: Scripps Research Institute; 2004.
 26. Wang J, et al. Development and testing of a general amber force field. *J Comput Chem.* 2004;25:1157–74.
 27. Cornell WD, et al. A second generation force field for the simulation of proteins, nucleic acids, and organic molecules. *J Am Chem Soc.* 1995;117:5179–97.
 28. Cerutti DS, et al. Staggered Mesh Ewald: an extension of the Smooth Particle-Mesh Ewald method adding great versatility. *J Chem Theory Comput.* 2009;5:2322.
 29. Simmerling C, et al. Combined locally enhanced sampling and Particle Mesh Ewald as a strategy to locate the experimental structure of a non-helical nucleic acid. *J Am Chem Soc.* 1998;120:7149–55.
 30. Gohlke H, et al. Insights into protein-protein binding by binding free energy calculation and free energy decomposition for the Ras-Raf and Ras-RalGDS complexes. *J Mol Biol.* 2003; 330:891–913.
 31. Kollman PA, et al. Calculating structures and free energies of complex molecules: combining molecular mechanics and continuum models. *Acc Chem Res.* 2000;33:889–97.
 32. Gund P. Three-dimensional pharmacophoric pattern searching. *Prog Mol Subcell Biol.* 1977;5:17.
 33. Guner OF. Pharmacophore perception, development, and use in drug design. La Jolla: International University Line; 2000.
 34. Langer T, Hoffmann RD. Pharmacophores and pharmacophore searches. Wiley: Weinheim; 2006.
 35. Wolber G, Langer T. LigandScout: 3-D pharmacophores derived from protein-bound ligands and their use as virtual screening filters. *J Chem Inf Model.* 2004;45:160–9.
 36. Friesner RA, et al. Glide: a new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy. *J Med Chem.* 2004;47:1739–49.
 37. Long J-F, et al. Supramodular structure and synergistic target binding of the N-terminal tandem PDZ domains of PSD-95. *J Mol Biol.* 2003;327:203–14.
 38. Feng W, et al. PDZ7 of glutamate receptor interacting protein binds to its target via a novel hydrophobic surface area. *J Biol Chem.* 2002; 277:41140–6.



<http://www.springer.com/978-1-62703-007-6>

Rational Drug Design

Methods and Protocols

Zheng, Y. (Ed.)

2012, XI, 230 p. 58 illus., 22 illus. in color., Hardcover

ISBN: 978-1-62703-007-6

A product of Humana Press