

# Metal Transporters in Plants

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**Abstract** Several transition metals are essential for plants as for most other organisms. These elements have been needed in the course of evolution because of their chemical properties such as redox activity under physiological conditions (Cu, Fe) or Lewis acid strength (Zn). The properties that make transition metal ions indispensable for life, however, are also the reason why they can easily be toxic when present in excess. The main threat lies in their ability to produce reactive oxygen species (ROS). Unfortunately, toxic metals such as cadmium, lead, mercury, etc., as well as the essential ones can also produce ROS. In the course of industrialization, emissions of metals have risen tremendously and significantly exceed those from natural sources for practically all metals. Due to this mobilization of metals into the biosphere, their circulation through soil, water, and air has greatly increased. The main aim of this chapter is to discuss the effects of metal ions on a plant cell, to summarize the current state of the art in the field of thiol-rich compounds like phytochelatins to detoxify metal ions.

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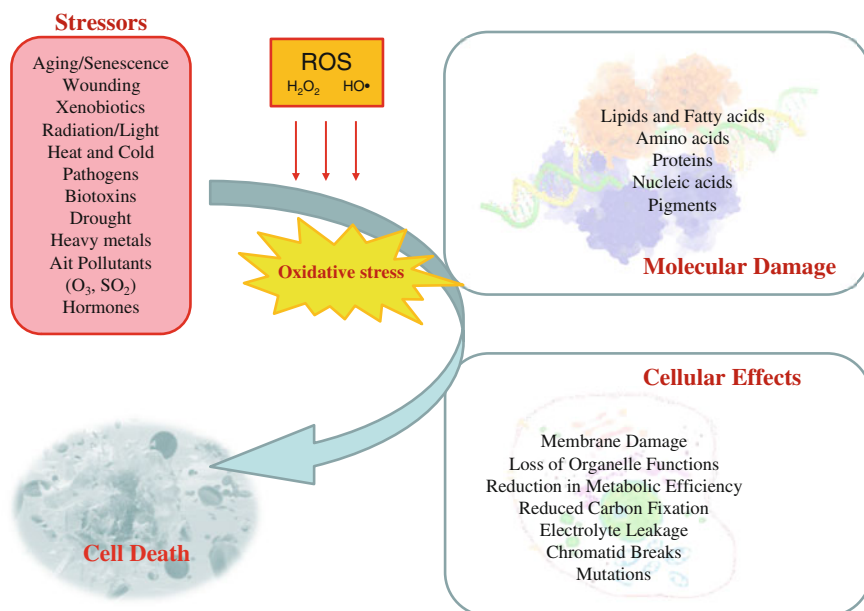
## Contents

1	Introduction.....	20
2	Plants and Heavy Metals.....	21
2.1	Glutathione and Related Thiols .....	23
2.2	Metallothionein-like Proteins and Metallothionein Expression .....	24
2.3	Induction of Thiols by Heavy Metals.....	24
3	Accumulation of Heavy Metals by Different Plant Species .....	25
3.1	Phytoremediation .....	25
3.2	Basic Mechanisms of Phytoremediation.....	26
3.3	Hyperaccumulator .....	27
4	Conclusion .....	31
	References.....	32

## 1 Introduction

Several transition metals are essential for plants as for most other organisms (Pilon et al. 2009; Puig and Penarrubia 2009). These elements have been acquired in the course of evolution because of their chemical properties such as redox activity under physiological conditions (Cu, Fe) or Lewis acid strength (Zn) (Welch 1995; Palmer and Guerinot 2009). The same properties that make transition metal ions indispensable for life, however, are also the reason why they can easily be toxic when present in excess. The main threat lies in their ability to produce reactive oxygen species (ROS) (Gratao et al. 2005). Unfortunately, toxic metals such as cadmium, lead, mercury, etc., as well as the essential ones can also produce ROS (Rodriguez-Serrano et al. 2009; Martins et al. 2011), see in Fig. 1.

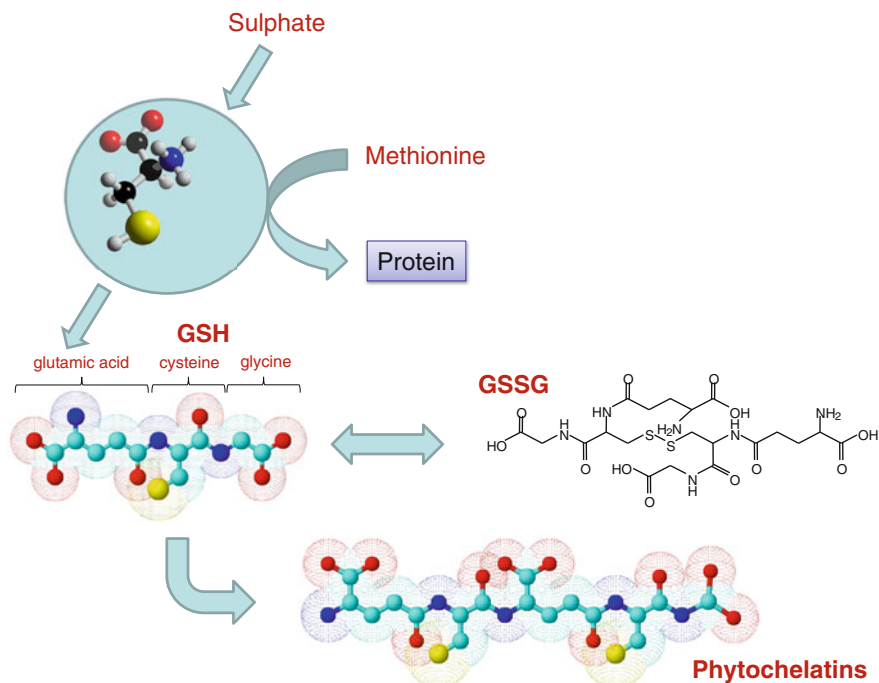
In the course of industrialization, emissions of metals have been tremendously raised and significantly exceed those from natural sources for practically all metals. Due to this mobilization of metals into the biosphere, their circulation through soil, water, and air has greatly increased (Kafka and Punccharova 2002; Cheng 2003; Boran and Altinok 2010; Yabe et al. 2010). Investigation of the influence of metals on an organism as well as of their transport and of maintaining their levels inside a cell is thus very topical. The main aim of this chapter is to discuss the effects of metal ions on a plant cell, to summarize the current state of the art in the field of thiol-rich compounds like phytochelatins to detoxify metal ions, and to review methods used for detection and determination of both metal ions and thiols (Fig. 2).



**Fig. 1** Scheme showing some of the initiators (stressors) of reactive oxygen species (ROS) and the biological consequences leading to a variety of physiological dysfunctions that can lead to cell death

## 2 Plants and Heavy Metals

Heavy metals represent a group of metallic elements of density higher than  $5 \text{ g/cm}^3$ . Some of them are essential for normal plant growth and development because they are integral parts of many enzymes and other proteins (Welch 1995; Grusak et al. 1999; Darrah and Staunton 2000; Shaul 2002; Kramer et al. 2007). However, elevated concentrations of both essential and non-essential heavy metals lead to symptoms of toxicity with growth and development processes affected. Heavy metal phytotoxicity may result from alterations of numerous physiological processes caused at cellular/molecular level by inactivating enzymes, blocking functional groups of metabolically important molecules, displacing or substituting for essential elements, and disrupting membrane integrity (Sergio et al. 2000; Rakhshaei et al. 2009; Douchiche et al. 2010a). A rather common consequence of heavy metal poisoning is the enhanced production of ROS due to interference with electron transport activities. This increase in ROS exposes cells to oxidative stress leading to lipid peroxidation, biological macromolecule deterioration, membrane dismantling, ion leakage, and DNA-strand cleavage (Fig. 1). Plants resort to a series of defense mechanisms that control uptake, accumulation, and translocation of these dangerous elements and detoxify them by excluding the free ionic forms from the cytoplasm. In addition, heavy metals can replace essential metallic element(s) with symptoms of



**Fig. 2** Consequence of synthesis of biologically active thiols from cysteine to phytochelatin

deficiency. Some of them, such as cadmium or lead, induce formation of reactive oxygen/nitrogen species-free radicals, which are responsible for damage of biomolecules including DNA (Deng et al. 2010b; Iannone et al. 2010; Liu et al. 2010b). However, plants have many detoxification and tolerance mechanisms that enable to survive in a polluted soil containing toxic levels of heavy metal/metals. These mechanisms include (i) establishment of symbiotic associations with soil microorganisms such as mycorrhiza that restrict movement of heavy metal ions and uptake by the plant (Lin et al. 2007; Amir et al. 2008; Arriagada et al. 2009; Iram et al. 2009), (ii) binding to the cell walls and eventually to root exudates (Douchiche et al. 2010b; Colzi et al. 2011; Lang and Wernitznig 2011), (iii) reduced influx through the plasma membrane (Courbot et al. 2007; Gonzalez-Mendoza and Zapata-Perez 2008; Xiao et al. 2008; Lang and Wernitznig 2011), (iv) chelation in the cytosol by various ligands such as phytochelatin and metallothioneins and further heavy metals compartmentalization in vacuole (Prasad 1995; Hall 2002; Hasan et al. 2009), (v) action of proteins connected with the stress caused by heavy metals (heat shock proteins) (Neumann et al. 1994; Wollgiehn and Neumann 1999). Plants with enhanced tolerance to heavy metal ions are able to survive, grow, and reproduce on polluted soils and are, therefore, usually connected with processes of soil decontamination and remediation (Ow 1996; Navari-Izzo and Quartacci 2001; Sonmez et al. 2008). As it will be discussed in Sect. 3, decontamination and remediation of

the polluted environment by using modern, non-destructive, and environment-friendly technologies is a topical theme to study. One of such technologies is called phytoremediation, which describes the treatment of environmental problems using plants (Jabeen et al. 2009; Kotrba et al. 2009; Karami and Shamsuddin 2010; Shao et al. 2010; Vamerali et al. 2010). Therefore, plant species with the best properties to grow and remediate the heavy metal polluted environment are intensively searched for. There are many promising plants usable in phytoremediation, especially in *Brassicaceae* family as *Thlaspi caerulescens*, *Thlaspi praecox*, *Thlaspi goesingense*, and *Arabidopsis halleri* (Gawronski and Gawronska 2007).

## 2.1 Glutathione and Related Thiols

Glutathione is one of the most significant thiol compounds occurring in the plant and animal kingdoms. It occurs in all living organisms—in prokaryotic as well as eukaryotic. GSH is redox buffer protecting the cytosol and other parts of cells against reactive oxygen radicals (ROS), which are induced by biotic and abiotic stress. In organisms, glutathione occurs in two forms, as reduced glutathione (GSH) and oxidized glutathione (GSSG). Both glutathione forms are strictly maintained ratios, whose disturbance is able to indicate stress elicited by various stress factors (Anderson 1998; Asensi et al. 1999; Garrido et al. 2010; Bielawski and Joy 1986). GSSG originates by formation of disulfide bond/linkage between two molecules of GSH, when two arisen  $H^+$  atoms participate in the ascorbate–glutathione cycle toward generated ROS elimination. Regressive GSSG molecule regeneration proceeds under GSH catalysis by reduction and oxidation of  $NADPH + H^+$  (Ogawa 2005; Paradiso et al. 2008), which is shown in Fig. 2. Its concentration varies in plants in the range from 0.1 to 10 mM (Meister and Anderson 1983).

The earliest reference to glutathione is from 1888, when its presence in yeasts was demonstrated. Glutathione structure was described as late as in 1935. In the 1960s, GSH was intensively studied because of its connection with human body liquids (McGovern et al. 1958; Manso and Wroblewski 1958; Pisciotta and Daly 1960a, b); though Dr. Alton Meister had indisputably the most contribution on glutathione metabolism clarification (Meister and Anderson 1983), which is proved by more than 4300 citations of his work on Web of Science server. GSH is a tripeptide containing  $\gamma$ -glutamyl-cysteinyl-glycine (Mullineaux and Rausch 2005).

In higher plants, GSH has many important functions that have crucial roles in maintenance of cellular redox homeostasis, and also participates in heavy metals and xenobiotics detoxification. In consequence to these functions, GSH is also used as a signal molecule in cells. GSH/GSSG couple reduction (redox) potential is not only influenced by reciprocal rate of GSH/GSSG, but also by changes in GSH synthesis as well as degradation (Schneider et al. 1992; Herschbach and Rennenberg 1994; Gelhaye et al. 2003; Rausch et al. 2007; Liedschulte et al. 2010).

## 2.2 Metallothionein-like Proteins and Metallothionein Expression

These proteins are polypeptides sharing low molecular mass, high cysteine content with absence of aromatic amino acids and histidine, high metal content, and abundance of CysXCys sequences where X is an amino acid other than cysteine (Suh et al. 1998; Liu et al. 2000; He et al. 2002; Lu et al. 2003). Metallothioneins (MT) with molecular weight varying from 2 to 16 kDa are subdivided into three classes based on their structure (Class I: polypeptides with a location of cysteines closely related to those in equine renal metallothioneins; Class II: polypeptides with locations of cysteine only distantly related to those in equine renal MT; Class III: atypical, non-translationally synthesized metal thiolate polypeptides (Liu et al. 2000). The metalloproteins have the ability to bind both physiological metal (Zn, Cu) as toxic (Cd, Pb, As) through thio group (–SH) of cysteine residues. The family of metallothionein-like proteins with a carboxy-terminal (further in text C-terminal) Gly was for the first time characterized in the yeast *Schizosaccharomyces pombe* exposed to cadmium (Plocke 1991). Shortly thereafter, a larger series within the same peptide family was found in several plants exposed to various heavy metals. Phytochelatins have a structural relationship to glutathione; the homologues related to homoglutathione were called homophytochelatins and those related to hydroxymethylglutathione were designated as hydroxymethylphytochelatins. Peptides having a C-terminal amino acid other than Gly are named as isophytochelatin with the parenthetic addition of the C-terminal amino acid. The prefix iso- was chosen to signify the equal called  $\gamma$ -GluCys peptides based on the common structural element. Specific thiols are named according to the sequence of amino acids (Grill et al. 1986; Rauser 1990; Cobbett 2000, 2001; Cobbett and Goldsbrough 2002; Pal and Rai 2010). There were also prepared plants carrying MT gene as a way to increase the ability of a plant to withstand metal ions (Diopan et al. 2008; Janouskova et al. 2005a, b; Kotrba et al. 1999; Macek et al. 2002, 1996; Pavlikova et al. 2004; Shestivska et al. 2011).

## 2.3 Induction of Thiols by Heavy Metals

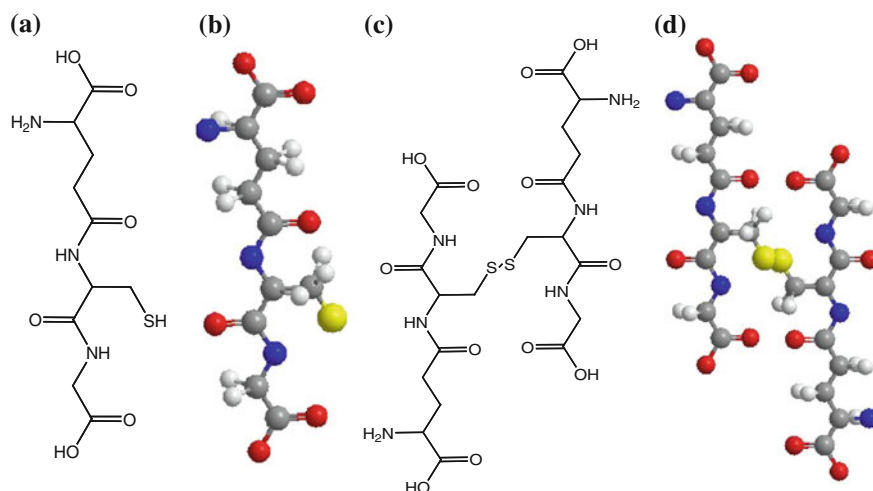
Various metals cause the appearance of thiols in plants (Kneer and Zenk 1992; Patra and Sharma 2000; Schmoger et al. 2000; Lee and Korban 2002; Abercrombie et al. 2008; Torres et al. 1997; Davis et al. 2006; Wang and Wang 2011). Induction of phytochelatins depends on the type of heavy metal as well as on the plant chosen. Silver through Cu (I) are class B metal ions that seek out nitrogen and sulfur centers in biological systems. Copper (II) and zinc are borderline metals that can form stable complexes with ligands offering oxygen, nitrogen, or sulfur atoms. Selenate, Te, and W are outside the grouping. All metals induced phytochelatins, except those members of the order of *Fabales* without

glutathione, where homophytochelatin were accompanied by the  $(\gamma\text{-GluCys})_n$  family except with Ni and Se (Tukendorf et al. 1997; Gupta et al. 2004; Loscos et al. 2006). Initial reports of induction by Ni, Se, Te, and W could not be repeated perhaps reflecting analytical problems at low phytochelatin levels. The abundance and length of induced  $\gamma\text{-GluCys}$  peptides varied with the used type of metal. In some cases, a somewhat effective metal at one concentration could not be tested at higher concentrations because its toxicity killed the cells. Cadmium is generally considered the most effective inducer of phytochelatin, but was surpassed by Ag in *Rubia tinctorum* (family *Rubiaceae*) root in vitro culture (Maitani et al. 1996). No phytochelatin induction was found for Al, Ca, Cr, Cs, K, Mg, Mn, Mo, Na, or V. Apart from Cr and Mn that are borderline metals, the above list of metals are all class A metals that share a strong preference for ligands with oxygen as the donor atom. Metals always inducing phytochelatin synthesis are Ag, Pb, Cd, and Zn. For details, see works focused on glutathione, metallothionein-like proteins and phytochelatin (Prasad 1995; di Toppo and Gabbriellini 1999; Zhang et al. 1999; Cobbett 2000; Clemens 2001; Cobbett and Goldsbrough 2002; Hall 2002; Pal et al. 2006; Mullainathan et al. 2007; Rausch et al. 2007; Ernst et al. 2008; Clemens and Persoh 2009; Jabeen et al. 2009; Yadav 2010; Hassinen et al. 2011).

### 3 Accumulation of Heavy Metals by Different Plant Species

#### 3.1 Phytoremediation

Phytoremediation is a widely accepted, aesthetically pleasant, solar-energy driven, passive technique that can be used to clean up sites with shallow, low to moderate levels of contamination (Padmavathiamma and Li 2007; Doran 2009; Jabeen et al. 2009; Memon and Schroder 2009; Zhao and McGrath 2009; Nwoko 2010). Phytoremediation is not only a growing science, it is also a growing industry. This technique can be used along with or, in some cases, in place of mechanical clean-up methods. Early estimates on the costs for remediating contaminated sites have shown that plants could do the same job as a group of engineers for one-tenth of the cost. The soil or water does not need to be gathered in and stored as hazardous waste, requiring large amounts of land, money, and manpower. Plants can be sown, watered, and then harvested with less manpower. The storage of the harvested plants as hazardous waste is seldom required and when needed it is less demanding than traditional disposal techniques. However, the main drawback of this novel technology is that it is not applicable to all sites. Several mechanisms may be involved in the direct and indirect action of phytoremediation in contaminated soils (Fig. 3).



**Fig. 3** Phytoremediation describes the treatment of environmental problems (bioremediation) using plants that mitigate the environmental problem without the need to excavate the contaminant material and dispose of it elsewhere. A range of processes mediated by plants or algae are useful in treating environmental problems as phytoextraction, phytostabilization, and (phyto) rhizofiltration. Phytoremediation consists of mitigating pollutant concentrations in contaminated soils, water, or air, with plants able to contain, degrade, or eliminate metals, pesticides, solvents, explosives, crude oil and its derivatives, and various other contaminants from the media that contain them

Therefore, phytoremediation of heavy metals can be divided into three groups:

1. Phytoextraction; the use of metal-accumulating plants to remove toxic metals from soil (Lasat 2002; McGrath and Zhao 2003; do Nascimento and Xing 2006; Van Nevel et al. 2007).
2. Phytostabilization; the use of plants to eliminate the bioavailability of toxic metals in soils (Cunningham and Berti 2000; Petrisor et al. 2004; Frerot et al. 2006; Kshirsagar and Aery 2007; Ehsan et al. 2009; Madejon et al. 2009; Andreazza et al. 2011).
3. (Phyto)rhizofiltration, the use of plant roots to remove toxic metals from polluted waters (Eapen et al. 2003; Verma et al. 2006; Khilji and Firdaus e 2008; Lee and Yang 2010; Yadav et al. 2011).

### 3.2 Basic Mechanisms of Phytoremediation

The remediation of soils contaminated with heavy metals is based on mechanisms of phytoextraction and phytostabilization. Phytoextraction, or phytoaccumulation, is referred to as the uptake and translocation of metal contaminants in the soil via the roots into the aboveground portions of the plants (Lasat 2002; McGrath and



Zhao 2003; do Nascimento and Xing 2006; Van Nevel et al. 2007). Risk element uptake by plants from soil depends on the level of pollution, forms of the element in soil, its mobility in the soil–plant system, and on plant species (Boruvka and Vacha 2006). Certain plants called hyperaccumulators absorb unusually large amounts of metals in comparison to other plants (e.g. up to 0.1 % chromium, cobalt, copper, or nickel or 1 % zinc, manganese in the aboveground shoots on a dry weight basis) (Xie et al. 2009; Masarovicova et al. 2010; Mengoni et al. 2010). Such hyperaccumulators are taxonomically widespread throughout the plant kingdom and are relevant to phytoremediation (Cunningham et al. 1995; Cunningham and Lee 1995). Phytoextraction is the using of hyperaccumulating plant species to remove metals from the soil by absorption into the roots and shoots of the plant. Metal concentrations in the shoots of some known hyperaccumulators can reach of extremely high levels (summarized by Cunningham and Ow 1996).

### 3.3 Hyperaccumulator

To physically remove metals from the contaminated site, the aboveground shoots of the hyperaccumulator plants are harvested and subsequently disposed of as hazardous wastes or treated for the recovery of the metals. Phytoremediation can be used to remove not only metals (e.g. Ag, Cd, Co, Cr, Cu, Hg, Mn, Mo, Ni, Pb, Zn) (Juwarkar et al. 2010) but also radionuclides (e.g.  $^{90}\text{Sr}$ ,  $^{137}\text{Cs}$ ,  $^{239}\text{Pu}$ ,  $^{234}\text{U}$ ,  $^{238}\text{U}$ ) (Cook et al. 2009; Fulekar et al. 2010; Hegazy and Emam 2010; Cerne et al. 2011) and certain organic compounds (i.e. petroleum hydrocarbons) (Abhilash et al. 2009; Gerhardt et al. 2009; Hussain et al. 2009; Perelo 2010; Megharaj et al. 2011). Plants growing in metal contaminated environments can accumulate toxic metal ions and efficiently compartmentalize them into various plant parts. Several studies indicated that the partitioning of heavy metals at the whole plant level can broadly be divided into three categories. For instance, Chaney and Giordano classified Mn, Zn, Cd, B, Mo, and Se as elements, which were readily translocated to the plant shoots; Ni, Co, and Cu, were intermediate, and Cr, Pb, and Hg were translocated to the lowest extent (Alloway 1995).

Jaffre et al. (1976) first applied the term hyperaccumulation to describe a highly abnormal level of metal accumulation in the title of their paper on nickel concentration in the tree *Sebertia acuminata* (family *Sapotaceae*) (Jaffre et al. 1976). The specific use of hyperaccumulation to denote a defined concentration (higher than  $1000 \text{ mg kg}^{-1}$  of  $\text{Ni kg}^{-1}$ ) was introduced by (Brooks et al. 1977) in discussing Ni concentration in species of *Homalium* (family *Flacoutiaceae*) and *Hybantus* (family *Violaceae*) from various parts of the world (Brooks et al. 1977). To date, more than 440 hyperaccumulator species have been described, three quarters of these being Ni accumulators from extensive occurrences of Ni-rich ultramafic soils found in many parts of the world (Martens and Boyd 1994; Borhidi 2001; Reeves 2006; Bani et al. 2009; Cecchi et al. 2010). Chaney (1983) first suggested the concept of using hyperaccumulator plants to accumulate high

quantities of metals in plant biomass to remove heavy metals from contaminated soils (Chaney 1983). In addition to the low cost, phytoaccumulation has several other important advantages over the traditional soil removal/replacement remediation methods. For example, it is in situ, preserves top soil, reduces the secondary waste stream, is environmentally sustainable, and the plant ash may have economic value (Garbisu and Alkorta 2001; Hetland et al. 2001). The main attraction of using hyperaccumulators for phytoremediation that remove and concentrate large amounts of a particular element is the possibility of employing species that remove and concentrate large amounts of a particular element from the soil without significant chemical intervention, other than classical application of fertilizers. It is important that the metal concentration in harvested plant tissue is greater than that in soil. To define, e.g., Ni hyperaccumulation more precisely, Reeves (2006) defined the hyperaccumulator as a plant that can accumulate a metal concentration of at least  $1000 \text{ mg kg}^{-1}$  in the dry matter of any above-ground tissue (Reeves 2006).

Baker and Brooks (1989) have reported about 400 metal-accumulating wild plants that accumulate high concentrations of heavy metals in their shoots (Baker and Brooks 1989). Natural hyperaccumulator plants often grow slowly and have low biomass yield. *T. caerulescens* (family *Brassicaceae*) was reported as a hyperaccumulator of cadmium and zinc (Plessl et al. 2010; Tuomainen et al. 2010; Leitenmaier and Kupper 2011; Leitenmaier et al. 2011; Liu et al. 2011). It can accumulate over 3 % of zinc and at the same time over 0.1 % of cadmium per dry biomass. The practical use of this plant for phytoremediation is restricted by its small biomass yield (Robinson et al. 1998). Metal hyperaccumulators are highly attractive model organisms as they have overcome major physiological bottlenecks limiting metal accumulation in biomass and metal tolerance.

There are two general approaches to phytoextraction: continuous and chemically enhanced phytoextraction (do Nascimento and Xing 2006; Nowack et al. 2006; Evangelou et al. 2007; Meers et al. 2008; Rajkumar et al. 2009; Saifullah et al. 2009; Rajkumar et al. 2010). The first approach uses naturally hyperaccumulating plants with the ability to accumulate an exceptionally high metal content in the shoots. Hyperaccumulating plants usually hyperaccumulate only a specific metal and metals that are primarily accumulated (Ni, Zn and Cu) are not among the most important environmental pollutants. No plant species has yet been found that demonstrates a wide spectrum of hyperaccumulation (Watanabe 1997). Hyperaccumulators are also mostly slow growing, low biomass-producing species, lacking good agronomic characteristics (Cunningham et al. 1995). There is no evidence that natural hyperaccumulator plants can access a less soluble and bio-available pool of metals in soil.

In non-hyperaccumulating plants, factors limiting their potential for phytoextraction include small root uptake and little root-to-shoot translocation of heavy metals. Chemically enhanced phytoextraction has been shown to overcome the above problems (Banaaraghi et al. 2010; Barrutia et al. 2010; de Araujo and do Nascimento 2010; Komarek et al. 2010; Zaier et al. 2010; Lomonte et al. 2011; Zhao et al. 2011). Common crop plants with high biomass can be triggered to

accumulate high amounts of low bioavailable metals, when their mobility in the soil and translocation from the roots to the green part of plants was enhanced by the addition of mobilizing agents when the crop had reached its maximum biomass. The feasibility of chemically enhanced phytoextraction has been primarily studied for Pb and chelating agents as soil additives; less attention has been given to other metals and radionuclides or their mixtures (Meers et al. 2005).

### 3.3.1 Nickel

About 360 species worldwide are known to act as Ni hyperaccumulators (Reeves 2006). The plant families most strongly represented are the *Brassicaceae*, *Euphorbiaceae*, *Asteraceae*, *Flacourtiaceae*, *Buxaceae*, and *Rubiaceae*. About 90 other species are from more than 30 families, distributed throughout the plant kingdom.

### 3.3.2 Zinc and Lead

The discovery of zinc accumulation in certain *Viola* and *Thlaspi* species in the nineteenth century was followed by other species with more than 10,000 mg kg<sup>-1</sup> Zn accumulation, notably *A. halleri*. This plant is one of the closest relatives of *A. thaliana*. It has colonized calamine soils, which are highly contaminated with Zn, Cd, Pb as a consequence of industrial activities. In addition, some populations have been reported to contain more than 100 µg g<sup>-1</sup> dry biomass Cd in their leaves. In hydropony, *A. halleri* has been shown to tolerate at least 30-fold higher Zn and 10-fold higher Cd concentrations in roots than *A. thaliana* can tolerate (Roosens et al. 2008).

Lead is present in most soils and rocks at concentrations below 50 mg kg<sup>-1</sup> and generally shows relatively low mobility in soils and in vegetation which typically have less than 10 mg kg<sup>-1</sup> Pb. In cases when Pb does enter the plant roots in larger concentrations from Pb-enriched soils, significant translocation to the upper parts of the plant is uncommon. Increased concentrations of Pb in aboveground tissues can be caused by entering of the metal bound with dust and fine soil particles directly to leaves through stomata.

### 3.3.3 Cadmium

Cadmium is a nonessential heavy metal widespread in our environment because of contamination by power stations, metal industries, and waste incineration. Toxicity to living cells occurs at very low concentrations, with suspected carcinogenic effects in humans. However, the biological effects of this metal and the mechanisms of its toxicity are not yet clearly understood (Suzuki et al. 2001). Cd is one of the increasingly frequent contaminants of agricultural soils, where it is usually

present at 0.1–0.2 mg kg<sup>-1</sup> but occasionally has been detected at much higher levels in some regions. Cadmium contamination in agricultural soils is due to either excessive phosphate fertilization, use of sewage sludge as a soil amendment, or due to naturally high background levels (de Borne et al. 1998). Cadmium has no essential function in plants and at high concentrations is toxic to plants and animals. Uptake of Cd by plant roots depends on the concentration, the oxidation state of this metal in solution, and on the physical–chemical characteristics of the soils such as pH content of clay, minerals, and organic matter (Brokbertold et al. 2011; Gao et al. 2011; Hou et al. 2011; Kovacik et al. 2011; Mleczek et al. 2011; Redjala et al. 2011). Few plant species have shown to accumulate more than 100 mg kg<sup>-1</sup> into their tissue (*T. caerulea* and *A. halleri*, both *Brassicaceae*) (Ozturk et al. 2003; Zhao et al. 2003; Ueno et al. 2004; Tolra et al. 2006; Liu et al. 2008). Recently, high accumulation abilities by *Salix* (*Salicaceae*) were shown (Kuzovkina et al. 2004; Tlustos et al. 2007; Mleczek et al. 2011).

### 3.3.4 Cobalt and Copper

Normal concentrations of Co and Cu in plants are in the ranges 0.03–2 and 5–25 mg kg<sup>-1</sup>, respectively. The tupelo or black gum of the southeastern United States (*Nyssa sylvatica*, *Cornaceae*) is remarkable in being able to accumulate as much as 845 mg kg<sup>-1</sup> Co from normal soils (McLeod and Ciravolo 2007). However, even on cobalt-enriched soils, such as those derived from ultramafic rocks, plant Cu rarely exceeds 20 mg kg<sup>-1</sup>.

Extensive screening of many sites of mining and smelting activity throughout Zaire, through plant and soil sample collections and analysis, identified 30 hyperaccumulators of cobalt and 32 of copper, with 12 species being common to the two lists (Homer et al. 1991; Keeling et al. 2003; Li et al. 2003; Faucon et al. 2007; Ghaderian et al. 2009; Wang et al. 2004; Wang and Zhong 2011). The Co and Cu accumulators have been found in more than dozen families. It can be mentioned that Co and Cu hyperaccumulators are not restricted only to metalliferous soils.

### 3.3.5 Manganese

Manganese is an essential element activating some of the enzymes involved in citric cycle (tricarboxylic acid cycle) and a central role of manganese cluster complexes in oxidation of water to oxygen has been reported. Toxic levels fall in the range 1000–12,000 mg kg<sup>-1</sup>, depending on the plant species. Some species have been found with 1000–5000 mg kg<sup>-1</sup> Mn on soils with manganese mineralization (more than 1 % Mn) and on soils with lower concentrations. Ultramafic soils may have 1000–5000 mg kg<sup>-1</sup>, which is not regarded as strongly abnormal. Most records of Mn hyperaccumulation come from these areas. Other hyperaccumulators were found on ultramafic soils in New Caledonia with concentrations around 1000 mg kg<sup>-1</sup> (Reeves 2006), in six plant species concentrations exceeded

10,000 mg kg<sup>-1</sup>, nine species had at least one specimen above this level. Mn hyperaccumulators can be found among *Apocynaceae*, *Celastraceae*, *Clusiaceae*, *Myrtaceae*, *Phytolaccaceae*, and *Proteaceae* families (Xue et al. 2004, 2006, 2007, 2009; Fernando et al. 2006, 2008; Min et al. 2007; Mizuno et al. 2008; Peng et al. 2008; Yang et al. 2008; Dou et al. 2009; Deng et al. 2010a; Liu et al. 2010a).

### 3.3.6 Selenium

Selenium is an essential element for animal and human health, with remarkably narrow range between levels required to prevent deficiency diseases and those producing symptoms of toxicity. Soil content is generally 0.01–2 mg kg<sup>-1</sup>. Se-rich soils can be found in the western part of United States, Ireland, Queensland, Colombia, and Venezuela. In plant dry matter, Se concentrations are generally below 1 mg kg<sup>-1</sup>, 0.01 mg kg<sup>-1</sup> in areas of low Se soils. Plants with more than 100 mg kg<sup>-1</sup> are considered as hyperaccumulators since normal levels in plants are below 2 mg kg<sup>-1</sup> (Reeves 2006). Plant genera in which extreme accumulation of Se can be found include *Astragalus* (*Fabaceae-Leguminosae*), *Stanleya* (*Brassicaceae*), *Haplopappus*, and *Machaeranthera* (*Asteraceae*) (Pickering et al. 2003; Cruz-Jimenez et al. 2004; Freeman et al. 2006; Galeas et al. 2007; Hung and Xie 2008; Freeman et al. 2009; Freeman and Banuelos 2011). Plants show a very wide variation in Se accumulation, as much as two orders of magnitude and even within a single locality.

Metal tolerant species and hyperaccumulators are a valuable and potentially useful biological resource which represent great potential for use in a variety of strategies for soil bioremediation, but some of them have been very rarely collected (Reeves 2006). There seems to be an urgent need for greater exploration of European metalliferous soils, so that more species of hyperaccumulating plants can be found and the distribution and rarity of these species can be better defined.

## 4 Conclusion

To consider whether the specific plant specie is able or not to remediate the polluted environment, not only heavy metals content in the plant tissues, but also the distribution of such metal ions in the tissues must be analyzed. Coupling of chromatographic technique for determination of heavy metal stress-induced plant peptides and spectrometric method for detection of spatial distribution of metals of interest seems to be suitable toward this aim.

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## References

- Abercrombie JM, Halfhill MD, Ranjan P, Rao MR, Saxton AM, Yuan JS, Stewart CN (2008) Transcriptional responses of *Arabidopsis thaliana* plants to As (V) stress. *BMC Plant Biol* 8:1–15
- Abhilash PC, Jamil S, Singh N (2009) Transgenic plants for enhanced biodegradation and phytoremediation of organic xenobiotics. *Biotechnol Adv* 27:474–488
- Alloway BJ (1995) Heavy metals in soils, 2nd edn. Blackie edn. The University of Reading U.K, Glasgow
- Amir H, Jasper DA, Abbott LK (2008) Tolerance and induction of tolerance to Ni of arbuscular mycorrhizal fungi from New Caledonian ultramafic soils. *Mycorrhiza* 19:1–6
- Anderson ME (1998) Glutathione: an overview of biosynthesis and modulation. *Chem Biol Interact* 112:1–14
- Andreazza R, Bortolon L, Pieniz S, Giacometti M, Roehrs DD, Lambais MR, Camargo FAO (2011) Potential phytoextraction and phytostabilization of perennial peanut on copper-contaminated vineyard soils and copper mining waste. *Biol Trace Elem Res* 143:1729–1739
- Arriagada C, Aranda E, Sampedro I, Garcia-Romera I, Ocampo JA (2009) Interactions of *Trametes versicolor*, *Coriolopsis rigida* and the arbuscular mycorrhizal fungus *Glomus deserticola* on the copper tolerance of *Eucalyptus globulus*. *Chemosphere* 77:273–278
- Asensi M, Sastre J, Pallardo FV, Lloret A, Lehner M, Garcia-de-la Asuncion J, Vina J (1999) Ratio of reduced to oxidized glutathione as indicator of oxidative stress status and DNA damage. *Oxi Antioxid* 299:267–276
- Baker AJM, Brooks RR (1989) Terrestrial higher plants which hyperaccumulate metallic elements—a review of their distribution, ecology and phytochemistry. *Biorecovery* 1:1–7
- Banaaraghi N, Hoodaji M, Afyuni M (2010) Use of EDTA and EDDS for enhanced *Zea mays*’ phytoextraction of heavy metals from a contaminated soil. *J Resid Sci Technol* 7:139–145
- Bani A, Echevarria G, Mullaj A, Reeves R, Morel JL, Sulce S (2009) Nickel hyperaccumulation by *Brassicaceae* in serpentine soils of Albania and Northwestern Greece. *Northeast Nat* 16:385–404
- Barrutia O, Garbisu C, Hernandez-Allica J, Garcia-Plazaola JI, Becerril JM (2010) Differences in EDTA-assisted metal phytoextraction between metalicolous and non-metallicolous accessions of *Rumex acetosa* L. *Environ Pollut* 158:1710–1715
- Bielawski W, Joy KW (1986) Reduced and oxidized glutathione and glutathione-reductase activity in tissues of *Pisum sativum*. *Planta* 169:267–272
- Boran M, Altinok I (2010) A review of heavy metals in water, sediment and living organisms in the black sea. *Turk J Fish Quat Sci* 10:565–572
- Borhidi A (2001) Phylogenetic trends in Ni-accumulating plants. *S Afr J Sci* 97:544–547
- Boruvka L, Vacha R (2006) Litavka river alluvium as a model area heavily polluted with potentially risk elements. In: Morel JL, Echevarria G, Goncharova N (eds) *Phytoremediation of Metal-Contaminated Soils*, vol 68. NATO Science Series IV Earth and Environmental Sciences. Springer, Dordrecht, pp 267–298
- Brokbartold M, Wischermann M, Marschner B (2011) Plant availability and uptake of lead, zinc, and cadmium in soils contaminated with anti-corrosion paint from pylons in comparison to heavy metal contaminated urban soils. *Water Air Soil Pollut* 223:199–213
- Brooks RR, Lee J, Reeves RD, Jaffre T (1977) Detection of nickeliferous rocks by analysis of herbarium specimens of indicator plants. *J Geochem Explor* 7:49–57
- Cecchi L, Gabbrielli R, Arnetoli M, Gonnelli C, Hasko A, Selvi F (2010) Evolutionary lineages of nickel hyperaccumulation and systematics in European Alysseae (*Brassicaceae*): evidence from nrDNA sequence data. *Ann Bot* 106:751–767
- Cerne M, Smadis B, Strok M (2011) Uptake of radionuclides by a common reed (*Phragmites australis* (Cav.) Trin. ex Steud.) grown in the vicinity of the former uranium mine at Zirovski vrh. *Nucl Eng Des* 24:1282–1286

- Chaney RL (1983) Plant uptake in inorganic waste constituents. In: Parr JF, Marsh PB, Kla JM (eds) Land treatment of hazardous wastes. Noyes Data Corp, Park Bridge
- Cheng SP (2003) Heavy metal pollution in China: origin, pattern and control. *Environ Sci Pollut Res* 10:192–198
- Clemens S (2001) Molecular mechanisms of plant metal tolerance and homeostasis. *Planta* 212:475–486
- Clemens S, Persoh D (2009) Multi-tasking phytochelatin synthases. *Plant Sci* 177:266–271
- Cobbett CS (2000) Phytochelatin biosynthesis and function in heavy-metal detoxification. *Curr Opin Plant Biol* 3:211–216
- Cobbett CS (2001) Heavy metal detoxification in plants: phytochelatin biosynthesis and function. *IUBMB Life* 51:183–188
- Cobbett C, Goldsbrough P (2002) Phytochelatins and metallothioneins: roles in heavy metal detoxification and homeostasis. *Annu Rev Plant Biol* 53:159–182
- Colzi I, Doumet S, Del Bubba M, Fornaini J, Arnetoli M, Gabbrielli R, Gonnelli C (2011) On the role of the cell wall in the phenomenon of copper tolerance in *Silene paradoxa* L. *Environ Exp Bot* 72:77–83
- Cook LL, Inouye RS, McGonigle TP (2009) Evaluation of four grasses for use in phytoremediation of Cs-contaminated arid land soil. *Plant Soil* 324:169–184
- Courbot M, Willems G, Motte P, Arvidsson S, Roosens N, Saumitou-Laprade P, Verbruggen N (2007) A major quantitative trait locus for cadmium tolerance in *Arabidopsis halleri* colocalizes with HMA4, a gene encoding a heavy metal ATPase. *Plant Physiol* 144:1052–1065
- Cruz-Jimenez G, Gardea-Torresdey JL, Peralta-Videa J, De La Rosa G (2004) *Larrea tridentata* and *Salvia roemeriana*: potential selenium hyperaccumulator desert plant species. *Abstr Pap Am Chem Soc* 227:U1054–U1054
- Cunningham SD, Berti WR (2000) Phytoextraction and phytostabilization: technical, economic, and regulatory considerations of the soil-lead issue. In: Ellen LK, Todd AA, Joel RC(eds) *Phytoremediation of contaminated soil and water*, vol 664, pp 359–376
- Cunningham SD, Lee CR (1995) Phytoremediation: plant-based remediation of contaminated soils and sediments. In: *Bioremediation: science and applications*. Sssa Special Publications, Madison, pp 145–156
- Cunningham SD, Ow DW (1996) Promises and prospects of phytoremediation. *Plant Physiol* 110:715–719
- Cunningham SD, Berti WR, Huang JWW (1995) Phytoremediation of contaminated soils. *Trends Biotechnol* 13:393–397
- Darrah PR, Staunton S (2000) A mathematical model of root uptake of cations incorporating root turnover, distribution within the plant, and recycling of absorbed species. *Eur J Soil Sci* 51:643–653
- Davis AK, Hildebrand M, Palenik B (2006) Gene expression induced by copper stress in the diatom *Thalassiosira pseudonana*. *Eukaryot Cell* 5:1157–1168
- de Araujo JDT, do Nascimento CWA (2010) Phytoextraction of lead from soil from a battery recycling site: the use of citric acid and NTA. *Water Air Soil Pollut* 211:113–120
- de Borne FD, Elmayan T, de Roton C, de Hys L, Tepfer M (1998) Cadmium partitioning in transgenic tobacco plants expressing a mammalian metallothionein gene. *Mol Breeding* 4:83–90
- Deng H, Li MS, Chen YX, Luo YP, Yu FM (2010a) A new discovered manganese hyperaccumulator-*Polygonum pubescens* Blume. *Fresenius Environ Bull* 19:94–99
- Deng XP, Xia Y, Hu W, Zhang HX, Shen ZG (2010b) Cadmium-induced oxidative damage and protective effects of N-acetyl-L-cysteine against cadmium toxicity in *Solanum nigrum* L. *J Hazard Mater* 180:722–729
- di Toppi LS, Gabbrielli R (1999) Response to cadmium in higher plants. *Environ Exp Bot* 41:105–130

- Diopan V, Shestivska V, Adam V, Macek T, Mackova M, Havel L, Kizek R (2008) Determination of content of metallothionein and low molecular mass stress peptides in transgenic tobacco plants. *Plant Cell Tissue Organ Cult* 94:291–298
- do Nascimento CWA, Xing BS (2006) Phytoextraction: a review on enhanced metal availability and plant accumulation. *Sci Agric* 63:299–311
- Doran PM (2009) Application of plant tissue cultures in phytoremediation research: incentives and limitations. *Biotechnol Bioeng* 103:60–76
- Dou CM, Fu XP, Chen XC, Shi JY, Chen YX (2009) Accumulation and detoxification of manganese in hyperaccumulator *Phytolacca americana*. *Plant Biol* 11:664–670
- Douchiche O, Driouich A, Morvan C (2010a) Spatial regulation of cell-wall structure in response to heavy metal stress: Cadmium-induced alteration of the methyl-esterification pattern of homogalacturonans. *Ann Bot* 105:481–491
- Douchiche O, Soret-Morvan O, Chaibi W, Morvan C, Paynel F (2010b) Characteristics of cadmium tolerance in 'Hermes' flax seedlings: contribution of cell walls. *Chemosphere* 81:1430–1436
- Eapen S, Suseelan KN, Tivarekar S, Kotwal SA, Mitra R (2003) Potential for rhizofiltration of uranium using hairy root cultures of *Brassica juncea* and *Chenopodium amaranticolor*. *Environ Res* 91:127–133
- Ehsan M, Santamaria-Delgado K, Vazquez-Alarcon A, Alderete-Chavez A, De la Cruz-Landero N, Jaen-Contreras D, Molumeli PA (2009) Phytostabilization of cadmium contaminated soils by *Lupinus uncinatus* Schldl. *Span J Agric Res* 7:390–397
- Ernst WHO, Krauss GJ, Verkleij JAC, Wesenberg D (2008) Interaction of heavy metals with the sulphur metabolism in angiosperms from an ecological point of view. *Plant Cell Environ* 31:123–143
- Evangeliou MWH, Ebel M, Schaeffer A (2007) Chelate assisted phytoextraction of heavy metals from soil: effect, mechanism, toxicity, and fate of chelating agents. *Chemosphere* 68:989–1003
- Faucon MP, Shutcha MN, Meerts P (2007) Revisiting copper and cobalt concentrations in supposed hyperaccumulators from SC Africa: influence of washing and metal concentrations in soil. *Plant Soil* 30:29–36
- Fernando DR, Batianoff GN, Baker AJ, Woodrow IE (2006) In vivo localization of manganese in the hyperaccumulator *Gossia bidwillii* (Benth.) N. Snow and Guymer (Myrtaceae) by cryo-SEM/EDAX. *Plant Cell Environ* 29:1012–1020
- Fernando DR, Marshall AT, Gouget B, Carriere M, Collins RN, Woodrow IE, Baker AJ (2008) Novel pattern of foliar metal distribution in a manganese hyperaccumulator. *Funct Plant Biol* 35:193–200
- Freeman JL, Banuelos GS (2011) Selection of salt and boron tolerant selenium hyperaccumulator *Stanleya pinnata* genotypes and characterization of Se phytoremediation from agricultural drainage sediments. *Environ Sci Technol* 45:9703–9710
- Freeman JL, Zhang LH, Marcus MA, Fakra S, McGrath SP, Pilon-Smits EAH (2006) Spatial imaging, speciation, and quantification of selenium in the hyperaccumulator plants *Astragalus bisulcatus* and *Stanleya pinnata*. *Plant Physiol* 142:124–134
- Freeman JL, Quinn CF, Lindblom SD, Klamper EM, Pilon-Smits EAH (2009) Selenium protects the hyperaccumulator *Stanleya Pinnata* against black-tailed prairie dog herbivory in native seleniferous habitats. *Am J Bot* 96:1075–1085
- Frerot H, Lefebvre C, Gruber W, Collin C, Dos Santos A, Escarre J (2006) Specific interactions between local metallicolous plants improve the phytostabilization of mine soils. *Plant Soil* 282:53–65
- Fulekar MH, Singh A, Thorat V, Kaushik CP, Eapen S (2010) Phytoremediation of (137) Cs from low level nuclear waste using *Catharanthus roseus*. *Indian J Pure Appl Phys* 48:516–519
- Galeas ML, Zhang LH, Freeman JL, Wegner M, Pilon-Smits EAH (2007) Seasonal fluctuations of selenium and sulfur accumulation in selenium hyperaccumulators and related nonaccumulators. *New Phytol* 173:517–525



- Gao XP, Flaten DN, Tenuta M, Grimmett MG, Gawalko EJ, Grant CA (2011) Soil solution dynamics and plant uptake of cadmium and zinc by durum wheat following phosphate fertilization. *Plant Soil* 338:423–434
- Garbisu C, Alkorta I (2001) Phytoextraction: a cost-effective plant-based technology for the removal of metals from the environment. *Bioresour Technol* 77:229–236
- Garrido T, Mendoza J, Riveros R, Saez L (2010) Acute and chronic effect of copper on levels of reduced and oxidized glutathione and nutrient uptake of tomato plants. *J Plant Nutr Soil Sci* 173:920–926
- Gawronski SW, Gawronska H (2007) Plant taxonomy for phytoremediation. In: *Advanced Science and Technology for Biological Decontamination of Sites Affected by Chemical and Radiological Nuclear Agents*. NATO Sci Ser IV Earth Environ Sci 75:79–88
- Gelhay E, Rouhier N, Jacquot JP (2003) Evidence for a subgroup of thioredoxin h that requires GSH/Grx for its reduction. *FEBS Lett* 555:443–448
- Gerhardt KE, Huang XD, Glick BR, Greenberg BM (2009) Phytoremediation and rhizoremediation of organic soil contaminants: potential and challenges. *Plant Sci* 176:20–30
- Ghaderian SM, Movahedi M, Ghasemi R (2009) Uptake and accumulation of cobalt by *Alyssum bracteatum*, an endemic Iranian Ni hyperaccumulator. *Northeast Nat* 16:131–138
- Gonzalez-Mendoza D, Zapata-Perez O (2008) Mechanism of plant tolerance to potentially toxic elements. *Bol Soc Bot Mex* 82:53–61
- Gratao PL, Polle A, Lea PJ, Azevedo RA (2005) Making the life of heavy metal-stressed plants a little easier. *Funct Plant Biol* 32:481–494
- Grill E, Gekeler W, Winnacker EL, Zenk HH (1986) Homo-phytochelatin are heavy metal-binding peptides of homo-glutathione containing fabales. *FEBS Lett* 205:47–50
- Grusak MA, Pearson JN, Marentes E (1999) The physiology of micronutrient homeostasis in field crops. *Field Crop Res* 60:41–56
- Gupta DK, Tohoyama H, Joho M, Inouhe M (2004) Changes in the levels of phytochelatin and related metal-binding peptides in chickpea seedlings exposed to arsenic and different heavy metal ions. *J Plant Res* 117:253–256
- Hall JL (2002) Cellular mechanisms for heavy metal detoxification and tolerance. *J Exp Bot* 53:1–11
- Hasan SA, Fariduddin Q, Ali B, Hayat S, Ahmad A (2009) Cadmium: toxicity and tolerance in plants. *J Environ Biol* 30:165–174
- Hassinen VH, Tervahauta AI, Schat H, Karenlampi SO (2011) Plant metallothioneins-Metal chelators with ROS scavenging activity? *Plant Biol* 13:225–232
- He HZ, Zhu CM, Lu T, Zhang RQ, Zhao NM, Liu JY (2002) Modeling the cysteine rich domain of plant metallothionein-like protein. *Acta Bot Sin* 44:1155–1159
- Hegazy AK, Emam MH (2010) Accumulation and soil-to-plant transfer of radionuclides in the Nile delta coastal black sand habitats. *Int J Phytorem* 13:140–155
- Herscbach C, Rennenberg H (1994) Influence of glutathione (GSH) on net uptake of sulfate and sulfate transport in tobacco plants. *J Exp Bot* 45:1069–1076
- Hetland MD, Gallagher JR, Daly DJ, Hassett DJ, Heebink LV (2001) Processing of plants used to phytoremediate lead-contaminated sites. In: *Phytoremediation, Wetlands, and Sediments*. *Bioremediat Ser* 6:129–136
- Homer FA, Morrison RS, Brooks RR, Clemens J, Reeves RD (1991) Comparative-studies of nickel, cobalt, and copper uptake by some nickel hyperaccumulators of the genus *Alyssum*. *Plant Soil* 138:195–205
- Hou LY, Shi WM, Wei WH, Shen H (2011) Cadmium uptake, translocation, and tolerance in AHA1OX *Arabidopsis thaliana*. *Biol Trace Elem Res* 139:228–240
- Hung CY, Xie JH (2008) Development of an efficient plant regeneration system for the selenium-hyperaccumulator *Astragalus racemosus* and the nonaccumulator *Astragalus canadensis*. *Hort Science* 43:2138–2142
- Hussain S, Siddique T, Arshad M, Saleem M (2009) Bioremediation and phytoremediation of pesticides: recent advances. *Crit Rev Environ Sci Technol* 39:843–907

- Iannone MF, Rosales EP, Groppa MD, Benavides MP (2010) Reactive oxygen species formation and cell death in catalase-deficient tobacco leaf disks exposed to cadmium. *Protoplasma* 245:15–27
- Iram S, Ahmad I, Javed B, Yaqoob S, Akhtar K, Kazmi MR, Badar uz Z (2009) Fungal tolerance to heavy metals. *Pak J Bot* 41:2583–2594
- Jabeen R, Ahmad A, Iqbal M (2009) Phytoremediation of heavy metals: physiological and molecular mechanisms. *Bot Rev* 75:339–364
- Jaffre T, Brooks RR, Lee J, Reeves RD (1976) *Sebertia acuminata*-Hyperaccumulator of nickel from New-Caledonia. *Science* 193:579–580
- Janouskova M, Pavlikova D, Macek T, Vosatka M (2005a) Arbuscular mycorrhiza decreases cadmium phytoextraction by transgenic tobacco with inserted metallothionein. *Plant Soil* 272:29–40
- Janouskova M, Pavlikova D, Macek T, Vosatka M (2005b) Influence of arbuscular mycorrhiza on the growth and cadmium uptake of tobacco with inserted metallothionein gene. *Appl Soil Ecol* 29:209–214
- Juwarkar AA, Singh SK, Mudhoo A (2010) A comprehensive overview of elements in bioremediation. *Rev Environ Sci Biotech* 9:215–288
- Kafka Z, Puncocharova J (2002) Toxicity of heavy metals in nature. *Chem Listy* 96:611–617
- Karami A, Shamsuddin ZH (2010) Phytoremediation of heavy metals with several efficiency enhancer methods. *Afr J Biotech* 9:3689–3698
- Keeling SM, Stewart RB, Anderson CWN, Robinson BH (2003) Nickel and cobalt phytoextraction by the hyperaccumulator *Berkheya coddii*: Implications for polymetallic phytomining and phytoremediation. *Int J Phytorem* 5:235–244
- Khilji S, Firdaus e B (2008) Rhizofiltration of heavy metals from the tannery sludge by the anchored hydrophyte, *Hydrocotyle umbellata* L. *Afr J Biotech* 7:3714–3720
- Kneer R, Zenk MH (1992) Phytochelatin protect plant enzymes from heavy-metal poisoning. *Phytochemistry* 31:2663–2667
- Komarek M, Vanek A, Mrnka L, Sudova R, Szakova J, Tejnecky V, Chrastny V (2010) Potential and drawbacks of EDDS-enhanced phytoextraction of copper from contaminated soils. *Environ Pollut* 158:2428–2438
- Kotrba P, Macek T, Ruml T (1999) Heavy metal-binding peptides and proteins in plants. A review. *Collect Czech Chem Commun* 64:1057–1086
- Kotrba P, Najmanova J, Macek T, Ruml T, Mackova M (2009) Genetically modified plants in phytoremediation of heavy metal and metalloid soil and sediment pollution. *Biotechnol Adv* 27:799–810
- Kovacik J, Klejdus B, Hedbavny J, Zon J (2011) Significance of phenols in cadmium and nickel uptake. *J Plant Physiol* 168:576–584
- Kramer U, Talke IN, Hanikenne M (2007) Transition metal transport. *FEBS Lett* 581:2263–2272
- Kshirsagar S, Aery NC (2007) Phytostabilization of mine waste: growth and physiological responses of *Vigna unguiculata* (L.) Walp. *J Environ Biol* 28:651–654
- Kuzovkina YA, Knee M, Quigley MF (2004) Cadmium and copper uptake and translocation in five Willow (*Salix* L.) species. *Int J Phytorem* 6:269–287
- Lang I, Wernitznig S (2011) Sequestration at the cell wall and plasma membrane facilitates zinc tolerance in the moss *Pohlia drummondii*. *Environ Exp Bot* 74:186–193
- Lasat MM (2002) Phytoextraction of toxic metals: a review of biological mechanisms. *J Environ Qual* 31:109–120
- Lee S, Korban SS (2002) Transcriptional regulation of *Arabidopsis thaliana* phytochelatin synthase (AtPCS1) by cadmium during early stages of plant development. *Planta* 215:689–693
- Lee M, Yang M (2010) Rhizofiltration using sunflower (*Helianthus annuus* L.) and bean (*Phaseolus vulgaris* L. var. vulgaris) to remediate uranium contaminated groundwater. *J Hazard Mater* 173:589–596

- Leitenmaier B, Kupper H (2011) Cadmium uptake and sequestration kinetics in individual leaf cell protoplasts of the Cd/Zn hyperaccumulator *Thlaspi caerulescens*. *Plant Cell Environ* 34:208–219
- Leitenmaier B, Witt A, Witzke A, Stemke A, Meyer-Klaucke W, Kroneck PMH, Kupper H (2011) Biochemical and biophysical characterisation yields insights into the mechanism of a Cd/Zn transporting ATPase purified from the hyperaccumulator plant *Thlaspi caerulescens*. *Biochim Biophys Acta-Biomembr* 1808:2591–2599
- Li YM, Chaney RL, Brewer EP, Angle JS, Nelkin J (2003) Phytoextraction of nickel and cobalt by hyperaccumulator *Alyssum* species grown on nickel-contaminated soils. *Environ Sci Technol* 37:1463–1468
- Liedschulte V, Wachter A, An ZG, Rausch T (2010) Exploiting plants for glutathione (GSH) production: uncoupling GSH synthesis from cellular controls results in unprecedented GSH accumulation. *Plant Biotechnol J* 8:807–820
- Lin AJ, Zhang XH, Wong MH, Ye ZH, Lou LQ, Wang YS, Zhu YG (2007) Increase of multi-metal tolerance of three leguminous plants by arbuscular mycorrhizal fungi colonization. *Environ Geochem Health* 29:473–481
- Liu JY, Lu T, Zhao NM (2000) Classification and nomenclature of plant metallothionein-like proteins based on their cysteine arrangement patterns. *Acta Bot Sin* 42:649–652
- Liu MQ, Yanai JT, Jiang RF, Zhang F, McGrath SP, Zhao FJ (2008) Does cadmium play a physiological role in the hyperaccumulator *Thlaspi caerulescens*? *Chemosphere* 71:1276–1283
- Liu P, Tang XM, Gong CF, Xu GD (2010a) Manganese tolerance and accumulation in six Mn hyperaccumulators or accumulators. *Plant Soil* 335:385–395
- Liu XM, Kim KE, Kim KC, Nguyen XC, Han HJ, Jung MS, Kim HS, Kim SH, Park HC, Yun DJ, Chung WS (2010b) Cadmium activates *Arabidopsis* MPK3 and MPK6 via accumulation of reactive oxygen species. *Phytochemistry* 71:614–618
- Liu GY, Zhang YX, Chai TY (2011) Phytochelatin synthase of *Thlaspi caerulescens* enhanced tolerance and accumulation of heavy metals when expressed in yeast and tobacco. *Plant Cell Rep* 30:1067–1076
- Lomonte C, Doronila A, Gregory D, Baker AJM, Kolev SD (2011) Chelate-assisted phytoextraction of mercury in biosolids. *Sci Total Environ* 409:2685–2692
- Loscos J, Naya L, Ramos J, Clemente MR, Matamoros MA, Becana M (2006) A reassessment of substrate specificity and activation of phytochelatin synthases from model plants by physiologically relevant metals. *Plant Physiol* 140:1213–1221
- Lu T, Liu JY, Zhang RQ, Zhao NM (2003) Modeling rice rgMT as a plant metallothionein-like protein by the distance geometry and homology methods. *Acta Bot Sin* 45:1297–1306
- Macek T, Mackova M, Truksa M, Cundy AS, Kotrba P, Yancey N, Schouten WH (1996) Preparation of transgenic tobacco with a yeast metallothionein combined with a polyhistidine tail. *Chem Listy* 90:690
- Macek T, Mackova M, Pavlikova D, Szakova J, Truksa M, Cundy S, Kotrba P, Yancey N, Schouten WH (2002) Accumulation of cadmium by transgenic tobacco. *Acta Biotechnol* 22:101–106
- Madejon P, Burgos P, Cabrera F, Madejon E (2009) Phytostabilization of amended soils polluted with trace elements using the Mediterranean shrub: *Rosmarinus officinalis*. *Int J Phytorem* 11:542–557
- Maitani T, Kubota H, Sato K, Yamada T (1996) The composition of metals bound to class III metallothionein (phytochelatin and its desglycyl peptide) induced by various metals in root cultures of *Rubia tinctorum*. *Plant Physiol* 110:1145–1150
- Manso C, Wroblewski F (1958) Glutathione reductase activity in blood and body fluids. *J Clin Invest* 37:214–218
- Martens SN, Boyd RS (1994) The ecological significance of nickel hyperaccumulation—a plant-chemical defense. *Oecologia* 98:379–384
- Martins LL, Mourato MP, Cardoso AI, Pinto AP, Mota AM, Goncalves MDS, de Varennes A (2011) Oxidative stress induced by cadmium in *Nicotiana tabacum* L.: effects on growth

- parameters, oxidative damage and antioxidant responses in different plant parts. *Acta Physiol Plant* 33:1375–1383
- Masarovicova E, Kralova K, Kummerova M (2010) Principles of classification of medicinal plants as hyperaccumulators or excluders. *Acta Physiol Plant* 32:823–829
- McGovern JJ, Isselbacher K, Rose PJ, Grossman MS (1958) Observations on the glutathione (GSH) stability of red blood cells. *Ama J Dis Child* 96:502
- McGrath SP, Zhao FJ (2003) Phytoextraction of metals and metalloids from contaminated soils. *Curr Opin Biotechnol* 14:277–282
- McLeod KW, Ciravolo TG (2007) Cobalt uptake by *Nyssa aquatica*, *N-sylvatica* var. *biflora*, and *Taxodium distichum* seedlings. *Wetlands* 27:40–43
- Meers E, Ruttens A, Hopgood MJ, Samson D, Tack FMG (2005) Comparison of EDTA and EDDS as potential soil amendments for enhanced phytoextraction of heavy metals. *Chemosphere* 58:1011–1022
- Meers E, Tack FMG, Van Slycken S, Ruttens A, Laing GD, Vangronsveld J, Verloo MG (2008) Chemically assisted phytoextraction: a review of potential soil amendments for increasing plant uptake of heavy metals. *Int J Phytorem* 10:390–414
- Megharaj M, Ramakrishnan B, Venkateswarlu K, Sethunathan N, Naidu R (2011) Bioremediation approaches for organic pollutants: a critical perspective. *Environ Int* 37:1362–1375
- Meister A, Anderson ME (1983) Glutathione. *Annu Rev Biochem* 52:711–760
- Memon AR, Schroder P (2009) Implications of metal accumulation mechanisms to phytoremediation. *Environ Sci Pollut Res* 16:162–175
- Mengoni A, Schat H, Vangronsveld J (2010) Plants as extreme environments? Ni-resistant bacteria and Ni-hyperaccumulators of serpentine flora. *Plant Soil* 331(1–2):5–16
- Min Y, Tie BQ, Tang MZ, Aoyama I (2007) Accumulation and uptake of manganese in a hyperaccumulator *Phytolacca americana*. *Miner Eng* 20:188–190
- Mizuno T, Hirano K, Kato S, Obata H (2008) Cloning of ZIP family metal transporter genes from the manganese hyperaccumulator plant *Chengiopanax sciadophylloides*, and its metal transport and resistance abilities in yeast. *Soil Sci Plant Nutr* 54:86–94
- Mleczeck M, Kozłowska M, Kaczmarek Z, Magdziak Z, Golinski P (2011) Cadmium and lead uptake by *Salix viminalis* under modified Ca/Mg ratio. *Ecotoxicology* 20:158–165
- Mullainathan L, Arulbalachandran D, Lakshmanan GMA, Velu S (2007) Phytoremediation: metallophytes an effective tool to remove soil toxic metal. *Plant Arch* 7:19–23
- Mullineaux PM, Rausch T (2005) Glutathione, photosynthesis and the redox regulation of stress-responsive gene expression. *Photosynth Res* 86:459–474
- Navari-Izzo F, Quartacci MF (2001) Phytoremediation of metals-Tolerance mechanisms against oxidative stress. *Minerva Biotechnol* 13:73–83
- Neumann D, Lichtenberger O, Gunther D, Tschiersch K, Nover L (1994) Heat-shock proteins induce heavy-metal tolerance in higher-plants. *Planta* 194:360–367
- Nowack B, Schulin R, Robinson BH (2006) Critical assessment of chelant-enhanced metal phytoextraction. *Environ Sci Technol* 40:5225–5232
- Nwoko CO (2010) Trends in phytoremediation of toxic elemental and organic pollutants. *Afr J Biotech* 9:6010–6016
- Ogawa K (2005) Glutathione-associated regulation of plant growth and stress responses. *Antioxid Redox Signal* 7:973–981
- Ow DW (1996) Heavy metal tolerance genes: Prospective tools for bioremediation. *Resour Conserv Recycl* 18:135–149
- Ozturk L, Karanlik S, Ozkutlu F, Cakmak I, Kochian LV (2003) Shoot biomass and zinc/cadmium uptake for hyperaccumulator and non-accumulator *Thlaspi* species in response to growth on a zincdeficient calcareous soil. *Plant Sci* 164:1095–1101
- Padmavathiamma PK, Li LY (2007) Phytoremediation technology: hyper-accumulation metals in plants. *Water Air Soil Pollut* 184:105–126
- Pal R, Rai JPN (2010) Phytochelatin: peptides involved in heavy metal detoxification. *Appl Biochem Biotechnol* 160:945–963

- Pal M, Horvath E, Janda T, Paldi E, Szalai G (2006) Physiological changes and defense mechanisms induced by cadmium stress in maize. *J Plant Nutr Soil Sci* 169:239–246
- Palmer CM, Gueriot ML (2009) Facing the challenges of Cu, Fe and Zn homeostasis in plants. *Nat Chem Biol* 5:333–340
- Paradiso A, Berardino R, de Pinto MC, di Toppi LS, Storelli MM, Tommasi F, De Gara L (2008) Increase in ascorbate-glutathione metabolism as local and precocious systemic responses induced by cadmium in durum wheat plants. *Plant Cell Physiol* 49:362–374
- Patra M, Sharma A (2000) Mercury toxicity in plants. *Bot Rev* 66:379–422
- Pavlikova D, Macek T, Mackova M, Sura M, Szakova J, Tlustos P (2004) The evaluation of cadmium, zinc and nickel accumulation ability of transgenic tobacco bearing different transgenes. *Plant Soil Environ* 50:513–517
- Peng KJ, Luo CL, You WX, Lian CL, Li XD, Shen ZG (2008) Manganese uptake and interactions with cadmium in the hyperaccumulator-*Phytolacca americana* L. *J Hazard Mater* 154:674–681
- Perelo LW (2010) Review: *In situ* and bioremediation of organic pollutants in aquatic sediments. *J Hazard Mater* 177:81–89
- Petrisor IG, Dobrota S, Komnitsas K, Lazar I, Kuperberg JM, Serban M (2004) Artificial inoculation-Perspectives in tailings phytostabilization. *Int J Phytorem* 6:1–15
- Pickering IJ, Wright C, Bubner B, Ellis D, Persans MW, Yu EY, George GN, Prince RC, Salt DE (2003) Chemical form and distribution of selenium and sulfur in the selenium hyperaccumulator *Astragalus bisulcatus*. *Plant Physiol* 131:1460–1467
- Pilon M, Cohu CM, Ravet K, Abdel-Ghany SE, Gaymard F (2009) Essential transition metal homeostasis in plants. *Curr Opin Plant Biol* 12:347–357
- Pisciotta AV, Daly M (1960a) Reduced glutathione (GSH) content of leukocytes in various hematologic diseases. *Blood* 15:421–422
- Pisciotta AV, Daly M (1960b) Studies on Agranulocytosis.3. the reduced glutathione (GSH) content of leukocytes of normals and patients recovered from agranulocytosis. *Blood* 16:1572–1578
- Plessl M, Rigola D, Hassinen VH, Tervahauta A, Karenlampi S, Schat H, Aarts MGM, Ernst D (2010) Comparison of two ecotypes of the metal hyperaccumulator *Thlaspi caerulescens* (J. & C. PRESL) at the transcriptional level. *Protoplasma* 239:81–93
- Plocke DJ (1991) Cadmium-binding peptide complexes from *Schizosaccharomyces pombe*. *Methods Enzymol* 205:603–610
- Prasad MNV (1995) Cadmium toxicity and tolerance in vascular plants. *Environ Exp Bot* 35:525–545
- Puig S, Penarrubia L (2009) Placing metal micronutrients in context: transport and distribution in plants. *Curr Opin Plant Biol* 12:299–306
- Rajkumar M, Ae N, Freitas H (2009) Endophytic bacteria and their potential to enhance heavy metal phytoextraction. *Chemosphere* 77:153–160
- Rajkumar M, Ae N, Prasad MNV, Freitas H (2010) Potential of siderophore-producing bacteria for improving heavy metal phytoextraction. *Trends Biotechnol* 28:142–149
- Rakhshae R, Giah M, Pourahmad A (2009) Studying effect of cell wall's carboxyl-carboxylate ratio change of *Lemna minor* to remove heavy metals from aqueous solution. *J Hazard Mater* 163:165–173
- Rausser WE (1990) Phytochelatins. *Annu Rev Biochem* 59:61–86
- Rausch T, Gromes R, Liedschulte V, Muller I, Bogs J, Galovic V, Wachter A (2007) Novel insight into the regulation of GSH biosynthesis in higher plants. *Plant Biol* 9:565–572
- Redjala T, Zelko I, Sterckeman T, Legue V, Lux A (2011) Relationship between root structure and root cadmium uptake in maize. *Environ Exp Bot* 71:241–248
- Reeves R (2006) Hyperaccumulation of trace elements by plants. In: *Phytoremediation of metal-contaminated soils*. NATO Sci Ser IV Earth Environ Sci 68:25–52
- Robinson BH, Leblanc M, Petit D, Brooks RR, Kirkman JH, Gregg PEH (1998) The potential of *Thlaspi caerulescens* for phytoremediation of contaminated soils. *Plant Soil* 203:47–56

- Rodriguez-Serrano M, Romero-Puertas MC, Sparkes I, Hawes C, del Rio LA, Sandalio LM (2009) Peroxisome dynamics in *Arabidopsis* plants under oxidative stress induced by cadmium. *Free Radical Biol Med* 47:1632–1639
- Roosens N, Willems G, Saumitou-Laprade P (2008) Using *Arabidopsis* to explore zinc tolerance and hyperaccumulation. *Trends Plant Sci* 13(5):208–215
- Saifullah Meers E, Qadir M, de Caritat P, Tack FMG, Du Laing G, Zia MH (2009) EDTA-assisted Pb phytoextraction. *Chemosphere* 74:1279–1291
- Sergio C, Figueira R, Crespo AMV (2000) Observations of heavy metal accumulation in the cell walls of *Fontinalis antipyretica*, in a Portuguese stream affected by mine effluent. *J Bryol* 22:251–255
- Shao HB, Chu LY, Ruan CJ, Li H, Guo DG, Li WX (2010) Understanding molecular mechanisms for improving phytoremediation of heavy metal-contaminated soils. *Crit Rev Biotechnol* 30:23–30
- Shaul O (2002) Magnesium transport and function in plants: the tip of the iceberg. *Biometals* 15:309–323
- Shestivska V, Adam V, Prasek J, Macek T, Mackova M, Havel L, Diopan V, Zehnalek J, Hubalek J, Kizek R (2011) Investigation of the antioxidant properties of metallothionein in transgenic tobacco plants using voltammetry at a carbon paste electrode. *Int J Electrochem Sci* 6:2869–2883
- Schmoger MEV, Oven M, Grill E (2000) Detoxification of arsenic by phytochelatin in plants. *Plant Physiol* 122:793–801
- Schneider A, Martini N, Rennenberg H (1992) Reduced glutathione (GSH) transport into cultured tobacco cells. *Plant Physiol Biochem* 30:29–38
- Sonmez O, Bukun B, Kaya C, Aydemir S (2008) The assessment of tolerance to heavy metals (Cd, Pb and Zn) and their accumulation in three weed species. *Pak J Bot* 40:747–754
- Suh MC, Choi D, Liu JR (1998) Cadmium resistance in transgenic tobacco plants expressing the *Nicotiana glutinosa* L. metallothionein-like gene. *Mol Cells* 8:678–684
- Suzuki N, Koizumi N, Sano H (2001) Screening of cadmium-responsive genes in *Arabidopsis thaliana*. *Plant Cell Environ* 24:1177–1188
- Tlustos P, Szakova J, Vyslouzilova M, Pavlikova D, Weger J, Javorska H (2007) Variation in the uptake of arsenic, cadmium, lead, and zinc by different species of willows *Salix* spp. grown in contaminated soils. *Cent Eur J Biol* 2:254–275
- Tolra R, Pongrac P, Poschenrieder C, Vogel-Mikus K, Regvar M, Barcelo J (2006) Distinctive effects of cadmium on glucosinolate profiles in Cd hyperaccumulator *Thlaspi praecox* and non-hyperaccumulator *Thlaspi arvense*. *Plant Soil* 288:333–341
- Torres E, Cid A, Fidalgo P, Herrero C, Abalde J (1997) Long-chain class III metallothioneins as a mechanism of cadmium tolerance in the marine diatom *Phaeodactylum tricornutum* Bohlin. *Aquat Toxicol* 39:231–246
- Tukendorf A, SkorzynskaPolit E, Baszynski T (1997) Homophytochelatin accumulation in Cd-treated runner bean plants is related to their growth stage. *Plant Sci* 129:21–28
- Tuomainen M, Tervahauta A, Hassinen V, Schat H, Koistinen KM, Lehesranta S, Rantalainen K, Hayrinen J, Auriola S, Anttonen M, Karenlampi S (2010) Proteomics of *Thlaspi caerulescens* accessions and an inter-accession cross segregating for zinc accumulation. *J Exp Bot* 61:1075–1087
- Ueno D, Zhao FJ, Shen RF, Ma JF (2004) Cadmium and zinc accumulation by the hyperaccumulator *Thlaspi caerulescens* from soils enriched with insoluble metal compounds. *Soil Sci Plant Nutr* 50:511–515
- Vamerali T, Bandiera M, Mosca G (2010) Field crops for phytoremediation of metal-contaminated land. A review. *Environ Chem Lett* 8:1–17
- Van Nevel L, Mertens J, Oorts K, Verheyen K (2007) Phytoextraction of metals from soils: how far from practice? *Environ Pollut* 150:34–40
- Verma P, George KV, Singh HV, Singh SK, Juwarkar A, Singh RN (2006) Modeling rhizofiltration: heavy-metal uptake by plant roots. *Environ Model Assess* 11:387–394

- Wang MJ, Wang WX (2011) Cadmium sensitivity, uptake, subcellular distribution and thiol induction in a marine diatom: recovery from cadmium exposure. *Aquat Toxicol* 101:387–395
- Wang HO, Zhong GR (2011) Effect of organic ligands on accumulation of copper in hyperaccumulator and nonaccumulator *Commelina communis*. *Biol Trace Elem Res* 143:489–499
- Wang H, Shan XQ, Wen B, Zhang SZ, Wang ZJ (2004) Responses of antioxidative enzymes to accumulation of copper in a copper hyperaccumulator of *Commoelina communis*. *Arch Environ Contam Toxicol* 47:185–192
- Watanabe ME (1997) Phytoremediation on the brink of commercialization. *Environ Sci Technol* 31:A182–A186
- Welch RM (1995) Micronutrient nutrition of plants. *Crit Rev Plant Sci* 14:49–82
- Wollgiehn R, Neumann D (1999) Metal stress response and tolerance of cultured cells from *Silene vulgaris* and *Lycopersicon peruvianum*: role of heat stress proteins. *J Plant Physiol* 154:547–553
- Xiao S, Gao W, Chen QF, Ramalingam S, Chye ML (2008) Overexpression of membrane-associated acyl-CoA-binding protein ACBP1 enhances lead tolerance in *Arabidopsis*. *Plant J* 54:141–151
- Xie QE, Yan XL, Liao XY, Li X (2009) The arsenic hyperaccumulator fern *Pteris vittata* L. *Environ Sci Technol* 43:8488–8495
- Xu XG, Shi JY, Chen YX, Chen XC, Wang H, Perera A (2006) Distribution and mobility of manganese in the hyperaccumulator plant *Phytolacca acinosa* Roxb. (Phytolaccaceae). *Plant Soil* 285:323–331
- Xu XH, Chen XC, Shi JY, Chen YX, Wu WX, Perera A (2007) Effects of manganese on uptake and translocation of nutrients in a hyperaccumulator. *J Plant Nutr* 30:1737–1751
- Xu XH, Shi JY, Chen XC, Chen YX, Hu TD (2009) Chemical forms of manganese in the leaves of manganese hyperaccumulator *Phytolacca acinosa* Roxb. (Phytolaccaceae). *Plant Soil* 318:197–204
- Xue SG, Chen YX, Reeves RD, Baker AJM, Lin Q, Fernando DR (2004) Manganese uptake and accumulation by the hyperaccumulator plant *Phytolacca acinosa* Roxb. (Phytolaccaceae). *Environ Pollut* 131:393–399
- Yabe J, Ishizuka M, Umemura T (2010) Current levels of heavy metal pollution in Africa. *J Vet Med Sci* 72:1257–1263
- Yadav SK (2010) Heavy metals toxicity in plants: an overview on the role of glutathione and phytochelatins in heavy metal stress tolerance of plants. *S Afr J Bot* 76:167–179
- Yadav BK, Siebel MA, van Bruggen JJA (2011) Rhizofiltration of a heavy metal (lead) containing wastewater using the wetland plant *Carex pendula*. *Clean:Soil Air Water* 39:467–474
- Yang SX, Deng H, Li MS (2008) Manganese uptake and accumulation in a woody hyperaccumulator, *Schima superba*. *Plant Soil Environ* 54:441–446
- Zaier H, Ghnaya T, Ben Rejeb K, Lakhdar A, Rejeb S, Jemal F (2010) Effects of EDTA on phytoextraction of heavy metals (Zn, Mn and Pb) from sludge-amended soil with *Brassica napus*. *Bioresour Technol* 101:3978–3983
- Zhang YX, Chai TY, Burkard G (1999) Research advances on the mechanisms of heavy metal tolerance in plants. *Acta Bot Sin* 41:453–457
- Zhao FJ, McGrath SP (2009) Biofortification and phytoremediation. *Curr Opin Plant Biol* 12:373–380
- Zhao FJ, Lombi E, McGrath SP (2003) Assessing the potential for zinc and cadmium phytoremediation with the hyperaccumulator *Thlaspi caerulescens*. *Plant Soil* 249:37–43
- Zhao SL, Lian F, Duo LA (2011) EDTA-assisted phytoextraction of heavy metals by turfgrass from municipal solid waste compost using permeable barriers and associated potential leaching risk. *Bioresour Technol* 102:621–626

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